



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

An Interferon Regulated MicroRNA Provides Broad Cell-Intrinsic Antiviral Immunity through Multihit Host-Directed Targeting of the Sterol Pathway

Citation for published version:

Robertson, K, Hsieh, WY, Forster, T, Blanc, M, Lu, H, Crick, PJ, Yutuc, E, Watterson, S, Martin, K, Griffiths, S, Enright, AJ, Yamamoto, M, Madapura Marulasiddappa, P, Lennox, KA, Behlke, MA, Talbot, S, Haas, J, Dölken, L, Griffiths, WJ, Wang, Y, Angulo, A & Ghazal, P 2016, 'An Interferon Regulated MicroRNA Provides Broad Cell-Intrinsic Antiviral Immunity through Multihit Host-Directed Targeting of the Sterol Pathway', *PLoS Biology*, vol. 14, no. 3, e1002364. <https://doi.org/10.1371/journal.pbio.1002364>

Digital Object Identifier (DOI):

[10.1371/journal.pbio.1002364](https://doi.org/10.1371/journal.pbio.1002364)

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Publisher's PDF, also known as Version of record

Published In:

PLoS Biology

Publisher Rights Statement:

Copyright: © 2016 Robertson et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



RESEARCH ARTICLE

An Interferon Regulated MicroRNA Provides Broad Cell-Intrinsic Antiviral Immunity through Multihit Host-Directed Targeting of the Sterol Pathway

Kevin A. Robertson^{1,2*}, Wei Yuan Hsieh¹, Thorsten Forster^{1,2}, Mathieu Blanc^{1,3}, Hongjin Lu¹, Peter J. Crick⁴, Eylan Yutuc⁴, Steven Watterson^{1,5}, Kimberly Martin^{1,6}, Samantha J. Griffiths¹, Anton J. Enright⁷, Mami Yamamoto¹, Madapura M. Pradeepa⁸, Kimberly A. Lennox⁹, Mark A. Behlke⁹, Simon Talbot¹, Jürgen Haas¹, Lars Dölken^{10,11}, William J. Griffiths⁴, Yuqin Wang⁴, Ana Angulo^{12,13}, Peter Ghazal^{1,2*}



OPEN ACCESS

Citation: Robertson KA, Hsieh WY, Forster T, Blanc M, Lu H, Crick PJ, et al. (2016) An Interferon Regulated MicroRNA Provides Broad Cell-Intrinsic Antiviral Immunity through Multihit Host-Directed Targeting of the Sterol Pathway. *PLoS Biol* 14(3): e1002364. doi:10.1371/journal.pbio.1002364

Academic Editor: Hidde L Ploegh, Whitehead Institute, UNITED STATES

Received: May 28, 2015

Accepted: December 22, 2015

Published: March 3, 2016

Copyright: © 2016 Robertson et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: Time course microarray analysis data are available from the National Centre for Biotechnology Information Gene Expression Omnibus (GEO) under Super-Series accession number GSE42505 (Sub-Series numbers GSE63290). All other numeric data underlying the figures are available in the supplementary file [S1 Data](#).

Funding: The Centre for Systems Biology at Edinburgh is a Centre for Integrative Systems Biology (CISB) supported by the BBSRC and EPSRC. This work was supported by BBSRC (BB/K019112/1) to

1 Division of Pathway Medicine, University of Edinburgh, Edinburgh, United Kingdom, **2** SynthSys at Edinburgh University, The King's Buildings, Edinburgh, United Kingdom, **3** École Polytechnique Fédérale de Lausanne, Lausanne, Switzerland, **4** Institute of Mass Spectrometry, College of Medicine, Grove Building, Swansea University, Singleton Park, Swansea, United Kingdom, **5** Northern Ireland Centre for Stratified Medicine, University of Ulster, C-Tric, Altnagelvin Campus, Londonderry, Ireland, **6** Centre for Integrative Physiology, Edinburgh, United Kingdom, **7** EMBL – European Bioinformatics Institute, Wellcome Trust Genome Campus, Hinxton, Cambridge, United Kingdom, **8** The Institute of Genetics and Molecular Medicine, Western General Hospital, Edinburgh, United Kingdom, **9** Integrated DNA Technologies, Coralville, Iowa, United States of America, **10** Department of Medicine, University of Cambridge, Cambridge, United Kingdom, **11** Institute of Virology, University of Würzburg, Würzburg, Germany, **12** Institut d'Investigacions Biomèdiques August Pi i Sunyer, Barcelona, Spain, **13** Immunology Unit, Department of Cell Biology, Immunology, and Neurosciences, Medical School, University of Barcelona, Barcelona, Spain

* kevin.robertson@ed.ac.uk (KAR); p.ghazal@ed.ac.uk (PG)

Abstract

In invertebrates, small interfering RNAs are at the vanguard of cell-autonomous antiviral immunity. In contrast, antiviral mechanisms initiated by interferon (IFN) signaling predominate in mammals. Whilst mammalian IFN-induced miRNA are known to inhibit specific viruses, it is not known whether host-directed microRNAs, downstream of IFN-signaling, have a role in mediating broad antiviral resistance. By performing an integrative, systematic, global analysis of RNA turnover utilizing 4-thiouridine labeling of newly transcribed RNA and pri/pre-miRNA in IFN-activated macrophages, we identify a new post-transcriptional viral defense mechanism mediated by miR-342-5p. On the basis of ChIP and site-directed promoter mutagenesis experiments, we find the synthesis of miR-342-5p is coupled to the antiviral IFN response via the IFN-induced transcription factor, IRF1. Strikingly, we find miR-342-5p targets mevalonate-sterol biosynthesis using a multihit mechanism suppressing the pathway at different functional levels: transcriptionally via *SREBF2*, post-transcriptionally via miR-33, and enzymatically via *IDI1* and *SC4MOL*. Mass spectrometry-based lipidomics and enzymatic assays demonstrate the targeting mechanisms reduce intermediate sterol pathway metabolites and total cholesterol in macrophages. These results reveal a previously unrecognized mechanism by which IFN regulates the sterol pathway. The sterol pathway is known to be an integral part of the macrophage IFN antiviral response, and we

PG and WJG, BBSRC/EPSRC (BB/D019621/1) and the Wellcome Trust (WT066784/Z/02/Z) to PG and BBSRC sLoLa grant BB/K003801/1 to JH. The study was also supported by the Ministerio de Ciencia e Innovación (MICINN, Spain) through grant SAF 2014-55683 to AA. Reagent support was also provided by the RNAi Global Initiative. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: MAB and KAL are employed by Integrated DNA Technologies, Inc., (IDT) which offers oligonucleotides for sale similar to some of the compounds described in the manuscript. IDT is however not a publicly traded company and these authors do not personally own any shares/equity in IDT.

Abbreviations: BMDM, bone marrow-derived macrophage; ChIP, chromatin immunoprecipitation; CH25H, cholesterol 25-hydroxylase; DMAPP, isomer dimethylallyl diphosphate; EVL, ena-vasodilator stimulated phosphoprotein; FOH, farnesol; GGOH, geranylgeraniol; H β CD, (2-Hydroxypropyl)- β -cyclodextrin; HCMV, human cytomegalovirus; HSV1, Herpes Simplex virus 1; IFN, interferon; IFN- β , interferon beta; IFN- γ , interferon gamma; IFNAR1, interferon alpha receptor 1; IPP, isopentyl diphosphate; IRF, interferon regulatory factor; LXR, liver x receptor; MCMV, murine cytomegalovirus; MEV, mevalonolactone; MOI, multiplicity of infection; pMEF, primary murine embryo fibroblasts; Q-RT-PCR, quantitative reverse transcription polymerase chain reaction; siRNA, small interfering RNA; SQL, squalene; TYK2, tyrosine kinase 2; 25-HC, 25-hydroxycholesterol.

show that miR-342-5p exerts broad antiviral effects against multiple, unrelated pathogenic viruses such Cytomegalovirus and Influenza A (H1N1). Metabolic rescue experiments confirm the specificity of these effects and demonstrate that unrelated viruses have differential mevalonate and sterol pathway requirements for their replication. This study, therefore, advances the general concept of broad antiviral defense through multihit targeting of a single host pathway.

Author Summary

How infected cells respond to a virus during the first minutes to hours after infection can determine whether a disease develops and influences the host's long-term survival. In mammals, unlike plants and flies that use small RNAs to fight viral infections, virus-induced interferon responses are a critical early event resulting in broad protection against infection. Interferon is a secreted host protein that binds to receptors on the surface of infected and uninfected cells and activates biochemical pathways that profoundly change the expression of hundreds of cellular genes, including those encoding microRNAs. The antiviral functions of only a handful of these genes are understood, and it is not known how the majority contribute to broadly protect against many different viruses. In this study, we uncover an interferon-regulated microRNA (miR-342-5p) that contributes to broad host cell immunity against infection through the cholesterol biosynthesis pathway. We show that miR-342-5p does this through a multihit strategy, turning down the master regulator of sterol biosynthesis as well as several specifically targeted enzymes within the pathway. A wide range of viruses depend on a number of the metabolite side-branches of the sterol biosynthesis pathway for their replication. Notably, our study reveals that by utilising multihit targeting of key branch-points in a single pathway, miR-342-5p is able to inhibit the replication of unrelated, clinically significant pathogens ranging from Herpes to Flu viruses.

Introduction

The innate immune response plays a critical role in cellular resistance to infection. In plants and invertebrates, small interfering RNAs (siRNAs) play a vital role in cell-autonomous immunity to viruses. siRNAs are generated and amplified from viral RNA molecules by the cellular RNAi machinery and instil a profound protection against the respective pathogen [1,2]. In mammals, interferon (IFN)-mediated JAK-STAT signalling orchestrates the cellular response to infection and, for several decades, research has focused on the identification and characterisation of antiviral proteins [3]. In contrast, roles for small RNAs in infection of mammalian cells have yet to be fully established (Fig 1) [4].

Several cellular microRNAs have been shown to contribute positively or negatively to infection by targeting host or viral gene expression [5–8]. In this context, IFN stimulation alters the expression of hundreds of cellular genes including some microRNAs [6,9]. Importantly, however, antiviral functions have been attributed to very few of these, although evidence for IFN-induced miRNAs targeting specific pathogens has been documented [6]. A key unresolved issue with important therapeutic implications is: how can IFN-regulated miRNA enhance antiviral functionality? In 2007, Pedersen et al. showed that an IFN-elicited down-regulation of a virus-targeting host miRNA (miR-122) leads to a reduction in Hepatitis C virus replication. This was notable as, historically, studies of IFN antiviral effects have focused on induced genes

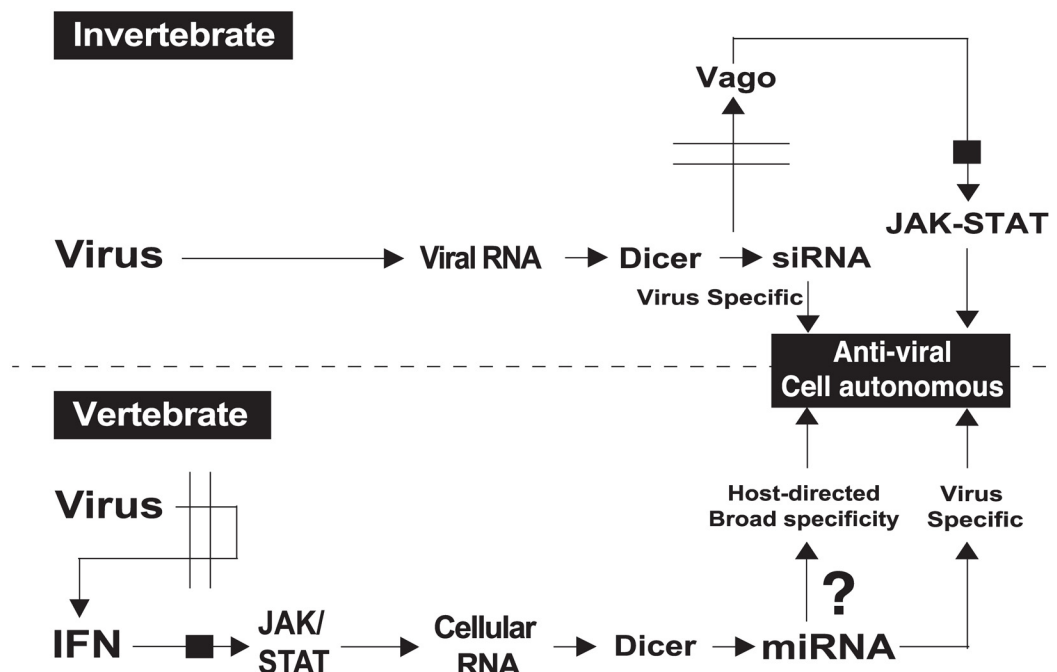


Fig 1. Schematic contrasting antiviral roles for small RNA and IFN-induced miRNA in invertebrates and vertebrates, respectively.

doi:10.1371/journal.pbio.1002364.g001

and this work, identifying a reduction in the expression of a virus-targeting host miRNA, has led to significant clinical advances [6,10]. Attention has also focused on interferon-regulated miRNA enhancement of IFN-mediated antiviral effects through the suppression of negative regulators such as the SOCS proteins [11]. Importantly, whilst antiviral miRNAs targeting host transcripts are emerging, few, if any, studies have characterised the precise mechanisms through which these function. miRNA targeting of multiple transcripts opens up potential for the simultaneous regulation of divergent and convergent cellular pathways and functions. It still remains an open question whether mammalian IFN-induced host-directed miRNAs act as effectors of the cell-intrinsic antiviral immune response. Such a strategy would arguably confer an advantage, as it has the potential to endow a broad-spectrum antiviral activity and greater resistance to the development of escape mutants.

Viruses rely on metabolic and biosynthetic cellular pathways for their replication, and a common feature is a dependency on cellular lipid metabolism [12]. As such, pharmacological inhibition of lipid biosynthesis can curtail virus replication [13–19]. Moreover, there is increasing evidence that the immune system and lipid pathways are tightly coupled [20–22] and defining how innate immunity and lipid metabolism are integrated, share resources, and crossregulate one-another during infection may result in new therapeutic strategies [20,23–26]. In this regard, IFN-induced inhibition of sterol biosynthesis serves as an integral component of the very early cellular response to virus infection and we, and others, have shown that cholesterol 25-hydroxylase (CH25H) and its cognate metabolite 25-hydroxycholesterol (25-HC) are important effectors in this response [18,24,27].

In this study, we sought to further unravel a previously unreported cellular mechanism underpinning immune regulation of lipid pathways and the inhibition of a broad range of viruses. We present evidence for a new miRNA-mediated cell-intrinsic antiviral effector arm of the IFN response, activated during the very early stages of infection. The miRNA under investigation has the capacity to instil a broad antiviral state in vitro and in vivo. Mechanistically, we

present data to show the broad antiviral functions of the miRNA occur via a specific multistep suppression of the sterol-biosynthetic pathway.

Results

IFN Uses Multiple Mechanisms to Suppress Sterol Biosynthesis

In experiments, seeking to address whether the activity of CH25H (UniProt: Q9Z0F5) was sufficient to account for the IFN-induced down-regulation of cholesterol biosynthesis in macrophages, we measured the abundance of several key sterol pathway transcripts including *HMGCS1* (Entrez Gene: 208715), *HMGCR* (Entrez Gene: 15357), *MVD* (Entrez Gene: 192156), *SQLE* (Entrez Gene: 20775), and *SREBF2* (Entrez Gene: 20788) in *CH25H*^{-/-} cells treated with interferon gamma (IFN- γ) (UniProt: P01580). Importantly, in the absence of CH25H, the effects of IFN- γ on these transcripts were reduced but not abrogated (Fig 2A and 2B) at 6 h (*HMGCR*, *SQLE* and *SREBF2*) and 24 h (*HMGCS1*, *HMGCR*, *MVD* and *SQLE*) after addition of the cytokine. These data suggested that 25-HC-dependent and independent mechanisms are involved early in the suppressive effect of IFN- γ on the sterol pathway.

To gain further kinetic insights into IFN- γ -elicited alterations in mRNA expression, we undertook a serial (every 30 mins for 8 h) high-resolution, systematic microarray analysis of de novo RNA synthesis and overall RNA abundance in bone marrow-derived macrophages (BMDM) treated with IFN- γ . A flow diagram summarising our protocol for the isolation and parallel analysis of newly transcribed and total RNA is presented (S1 Fig). These experiments revealed a significant down-regulation in the rate of transcript synthesis for 13/19 members of the cholesterol pathway (Data: Fig 3A and 3B and S2, S3 Figs Pathway: Fig 4) [28].

Decreases in the rate of pathway transcript synthesis commenced 2 h to 3 h after treatment (Fig 3A, S2 Fig) and were maintained for the duration of the time course (8 h). In addition, the rate of synthesis and abundance of *SREBF2* (but not *SREBF1* (Entrez Gene: 20787) also decreased in response to IFN- γ treatment (Fig 3C and 3D, S2 and S3 Figs). We note a more pronounced repression of *SREBF2* synthesis over abundance, which likely reflects an IFN- γ action on the well-characterised transcriptional autoregulation of SREBP2 [29]. Taken together, this data is consistent with previous studies [18,24,30,31], demonstrating that transcriptional mechanisms play a role in IFN- γ -mediated down-regulation of cholesterol. However, unexpected reductions in the abundance, but not synthesis of transcripts such as *MVD*, *NSDHL*, and *DHCR7* (Fig 3A and 3B, S2 and S3 Figs) suggested that cholesterol pathway transcripts may also be subject to 25-HC-independent post-transcriptional regulation. As IFN- γ -mediated suppression of the sterol pathway is strictly dependent on JAK-STAT signalling, we hypothesised that a likely post-transcriptional mechanism might involve IFN-stimulated miRNAs specifically targeting transcripts within the sterol metabolic network.

IFN Is Coupled to miR-342 Expression

In support of the above hypothesis, studies have documented miRNA which can regulate lipoprotein uptake (e.g., miR-125a and -455), lipid biosynthetic enzyme expression (e.g., miR-155, miR-21, and miR-185) and, in particular, cholesterol efflux (e.g., miR-33, miR-144) [32–37]. IFN- γ treatment of a melanoma cell line suggested that some of these miRNA (e.g., miR-125a, -455 and -185) may be part of an IFN response; however, it is not known if they are directly coupled to IFN [9]. Thus, we assessed changes in the expression of miRNA precursors (pri/pre-miRNAs) in BMDM stimulated with IFN- γ . Using conservative criteria for detection, we identified 66 pri/pre-miRNAs in our macrophage data (Fig 5A and 5B). Temporal analysis of our time course microarray data, however, revealed two of these, namely pri/pre-miR-155 (Entrez Gene: 387173) and pri/pre-miR-342 (Entrez Gene: 723909), to be significantly up-

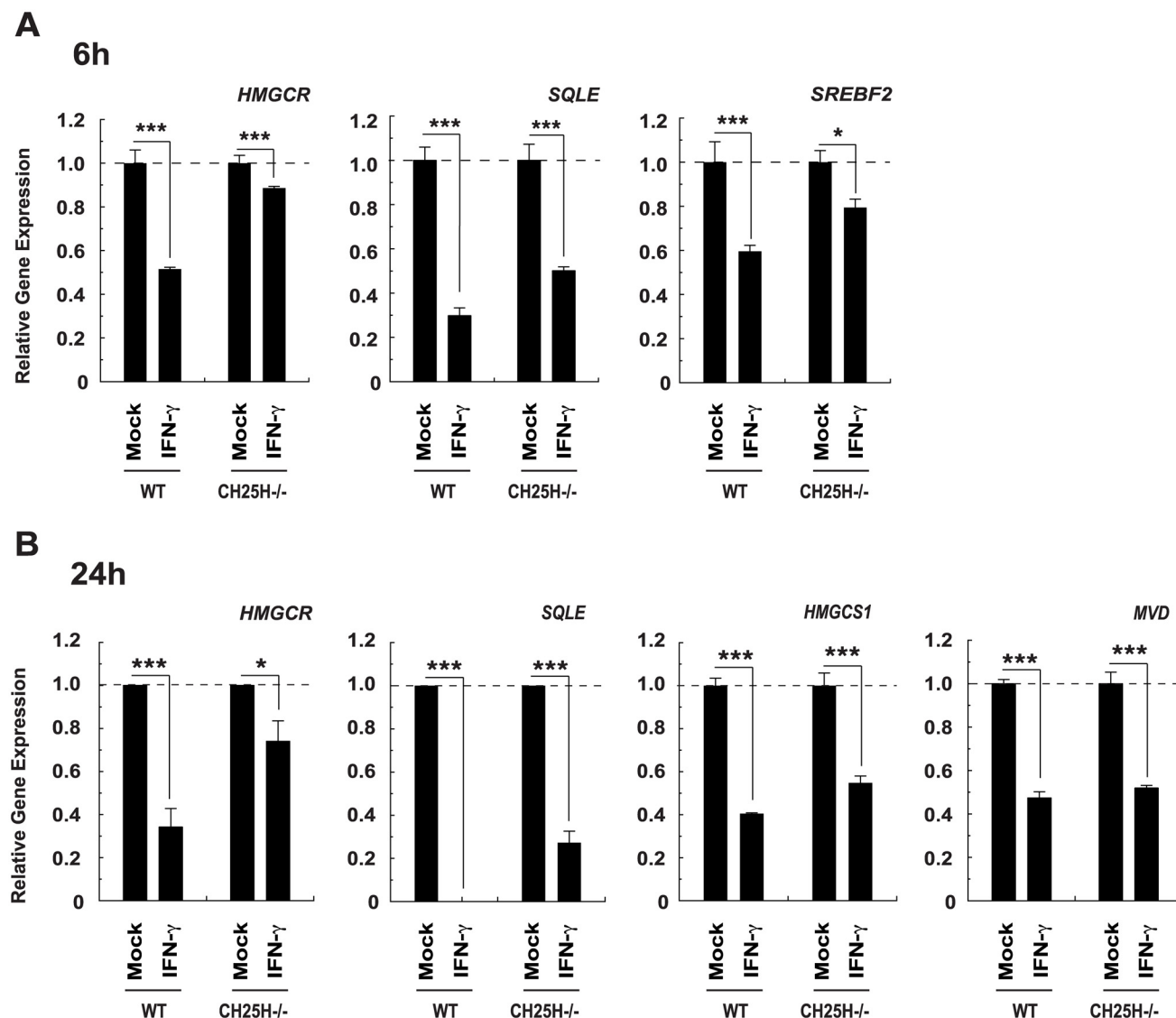


Fig 2. IFN- γ treatment of CH25H^{-/-} BMDM decreases the abundance of cholesterol biosynthesis pathway transcripts. (A) *HMGCR*, *SQLE*, and *SREBF2* transcript abundance in bone marrow-derive macrophage (BMDM) derived from wild-type or CH25H^{-/-} mice and treated with IFN- γ (10 U/ml) for 6 h. All data are mean \pm standard error of the mean (SEM). $n = 3$, *** $p \leq 0.001$ by 1-sample t test. (B) *HMGCR*, *SQLE*, *HMGCS1*, and *MVD* transcript abundance in BMDM derived from wild-type or CH25H^{-/-} mice and treated with IFN- γ (10 U/ml) for 24 h. All data are mean \pm SEM. *HMGCS1* data $n = 3$, *** $p \leq 0.001$ by 1-sample t test. *HMGCR* data $n = 9$, * $p \leq 0.05$, *** $p \leq 0.001$ by 1-sample t test. *MVD* data, $n = 3$, *** $p \leq 0.001$ by 1-sample t test. Squalene (SQL) data, wild-type BMDM $n = 3$: IFN- γ *** $p \leq 0.001$ by 1-sample t test, CH25H^{-/-} BMDM, $n = 6$, *** $p \leq 0.001$ by 1-sample t -test.

doi:10.1371/journal.pbio.1002364.g002

regulated during the first 8h of IFN- γ treatment (Fig 5A and 5B). MiR-155 is an evolutionarily highly conserved, NF- κ B-responsive miRNA encoded by the MIR155 host gene (MIR155HG). It is highly expressed in activated macrophages and lymphocytes [38]. Much less is known about the conserved miR-342 located in an intron of the Ena-vasodilator stimulated phospho-protein gene (*EVL*—Entrez Gene: 14026) in the mouse or Ena-Vasp-Like (*EVL*—Entrez Gene: 51466) in the human (Fig 6A) [39]. Gene expression data from 89 mouse cells and tissues analysed in the BioGPS GeneAtlas indicates the *EVL* transcript is primarily expressed in cells of the immune and nervous system [40]. In macrophages, miR-342 has previously been identified as a PU.1-regulated miRNA contributing to myeloid differentiation and a proinflammatory

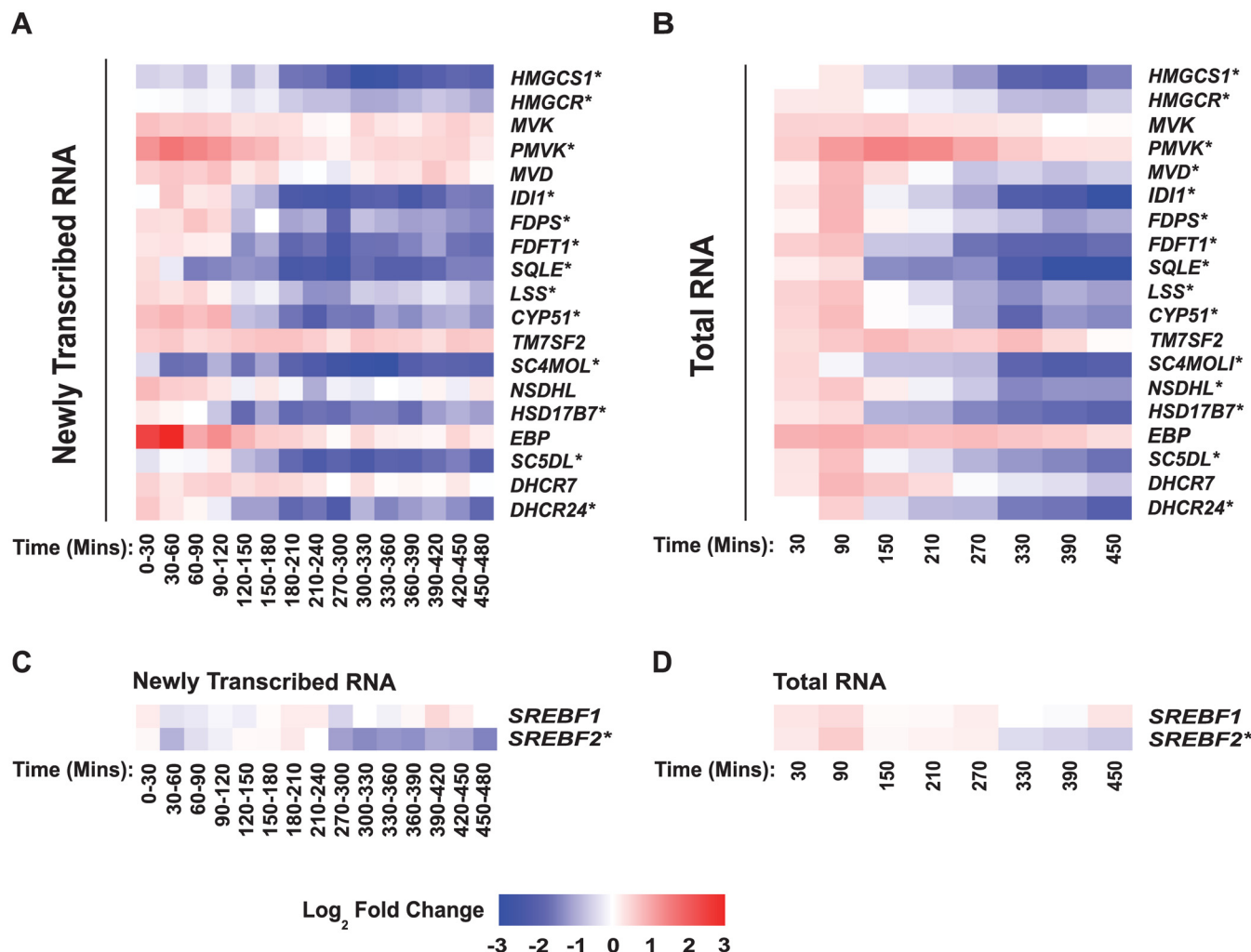


Fig 3. IFN- γ treatment of macrophages decreases the synthesis and abundance of cholesterol biosynthesis pathway transcripts. (A) Sequential analysis of sterol pathway-related transcript synthesis (every 30 min) in IFN- γ treated BMDM (relative to mock). (B) Abundance of sterol biosynthesis pathway-related transcripts in IFN- γ treated BMDM (relative to mock). (C) Synthesis of *SREBF1* and 2 transcripts in IFN- γ treated BMDM (relative to mock). (D) Abundance of *SREBF1* and 2 transcripts in IFN- γ treated BMDM (relative to mock). In Fig 3A, 3B, 3C, and 3D, each column represents a time period and each row one gene. Gene expression is shown as a pseudocolour: blue = decrease, red = increase. Log₂ fold change values were calculated by subtracting the Mock from the IFN- γ treated Log₂ scale signal value. * = Significant by MaSigPro test (see [Materials and Methods](#) and [S1 Methods](#)).

doi:10.1371/journal.pbio.1002364.g003

mediator capable of enhancing miR-155 expression [41,42]. To confirm the presence of mature miR-155, miR-342-3p and -5p derived from the precursors detected in our array analysis, we stimulated BMDM with IFN- γ (10 U/ml) or interferon beta (IFN- β) (Uniprot: P01575) (25 U/ml) and analysed miRNA expression using quantitative reverse transcription polymerase chain reaction (Q-RT-PCR). In these analyses, significant increases in the expression for all 3 miRNAs were observed (Fig 6B and 6C, S4A and S4B Fig).

Fig 6D and S4C Fig show the coordinate regulation of both the synthesis and abundance of the primary *EVL* transcript and pri/pre-miR-342 in IFN- γ stimulated BMDM. Both the primary transcript and precursor decreased in synthesis rate (relative to the mock) in the first 60 min after treatment. This was followed by an elevated synthesis rate and increased abundance from around 120 min onwards. Although the transcription factors IRF1 (UniProt: P10914) and IRF9 (UniProt: Q00978) have been implicated in the regulation of *EVL* in the context of

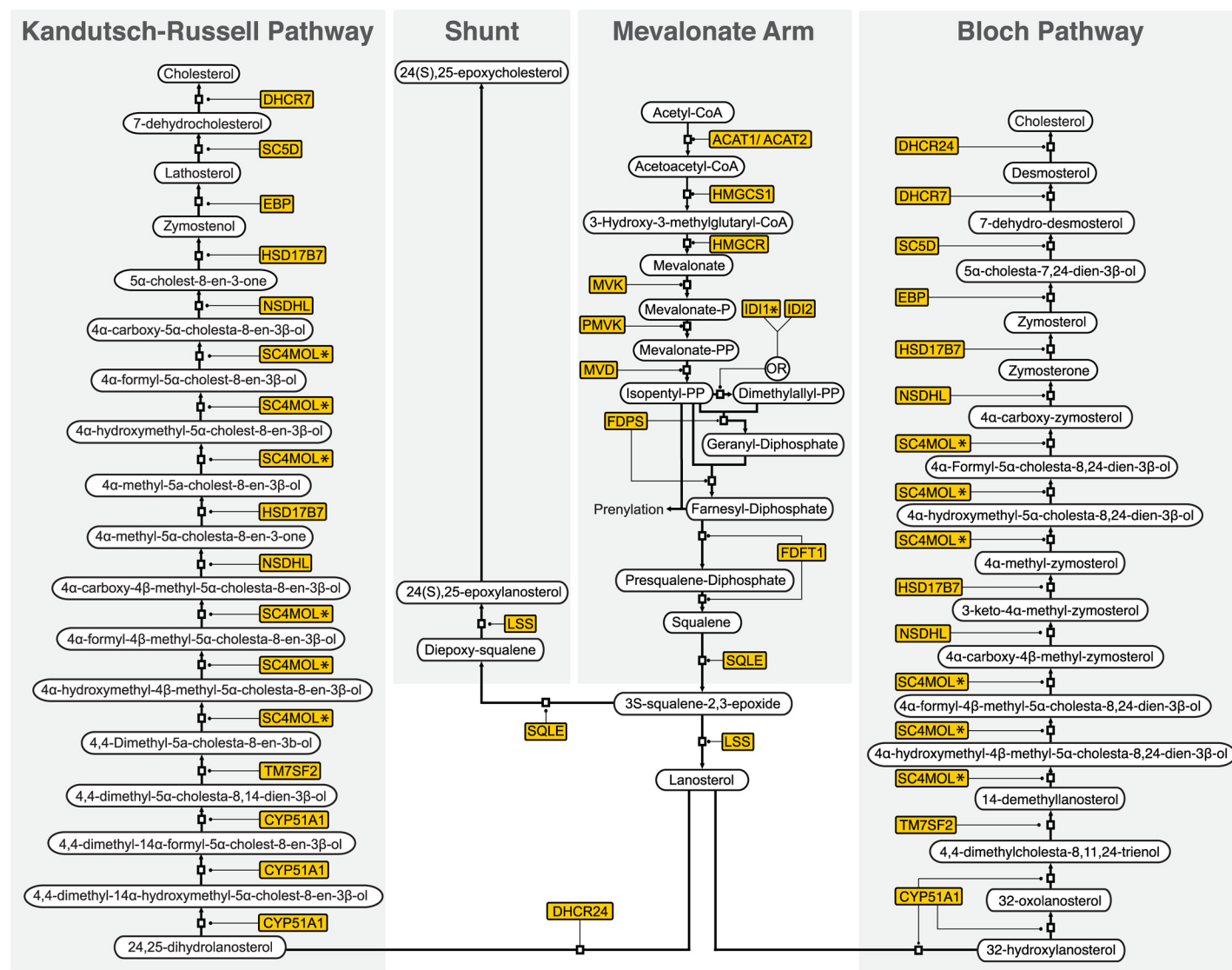


Fig 4. Schematic of the cholesterol biosynthesis pathway adapted from Mazein et al. (2013) [28]. * indicates pathway targets for miR-342-5p

doi:10.1371/journal.pbio.1002364.g004

long-term retinoic acid treatment of human promyelocytic leukaemia cells [43], it is not known whether the *EVL* promoter can be directly regulated by IFN and whether IFN signalling alone is sufficient for its induction [42]. To test the cell-autonomous requirement for IFN signalling in the up-regulation of *EVL* and pri/pre-miR-342 during infection, we analysed by microarray the endogenous regulation of these transcripts in BMDM with a genetic ablation in IFN beta (IFN- β), the type 1 IFN receptor (IFNAR1 –UniProt: P17181) (Fig 6E) or the downstream signalling molecule tyrosine kinase 2 (TYK2 UniProt: Q9R117) (S4D and S4E Fig). Ablation of IFN signalling abolished murine cytomegalovirus (MCMV)-induced induction of *EVL* and pri/pre-miR-342 in all cases demonstrating that virus-induced up-regulation of *EVL* and pri/pre-miR-342 is dependent on an intact type 1 IFN response. By computational promoter analysis, we identified three putative binding motifs (one IFN-stimulated response element/IRF1 binding site [ISRE/IRF1] and two potential IFN regulatory factor [IRF] binding sites) within a 1 kb region of the *EVL* promoter (S5A Fig). Luciferase reporter plasmids

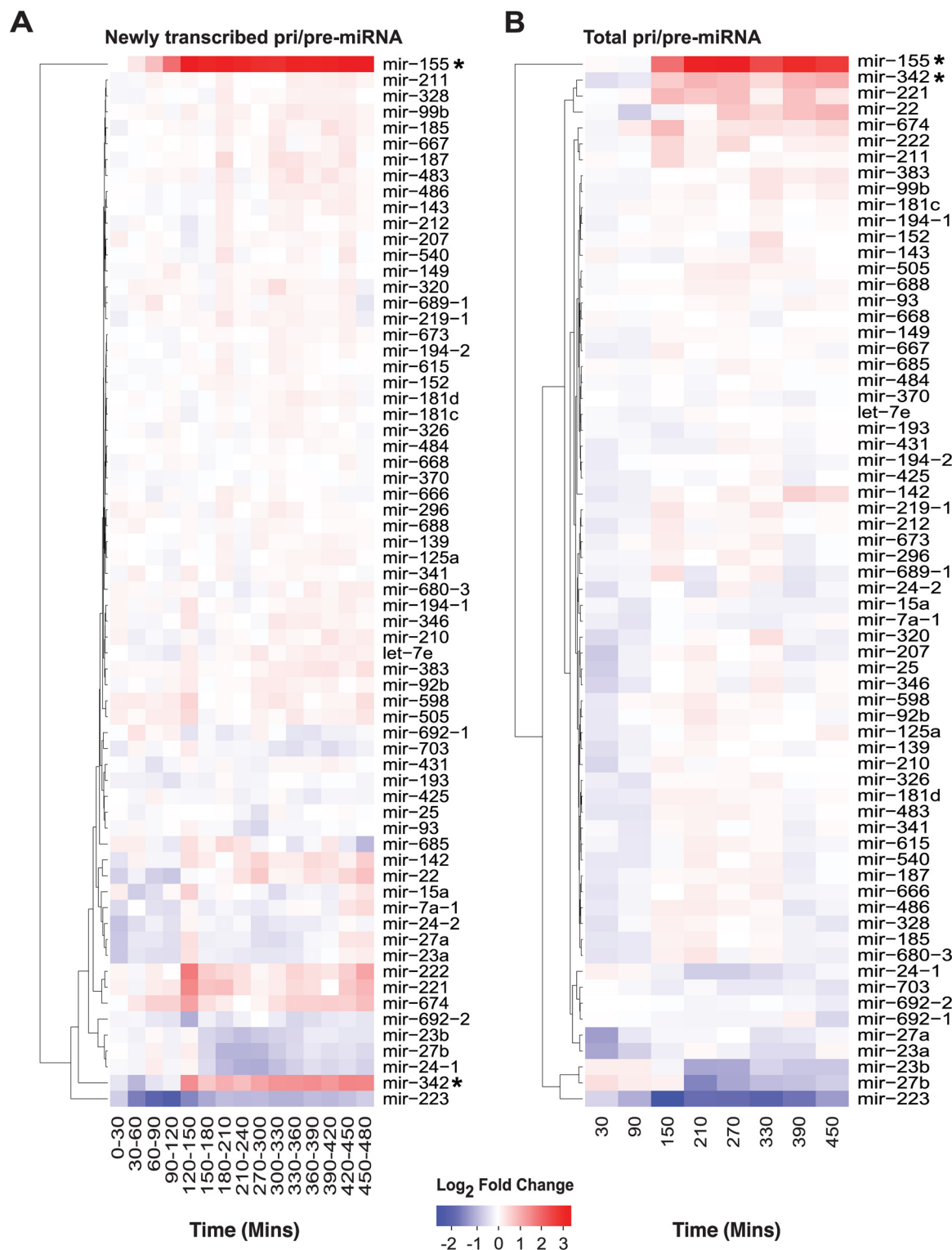


Fig 5. MiR-155 and miR-342 synthesis and abundance are significantly up-regulated in IFN- γ treated BMDM. (A and B) Pri/pre-miRNA synthesis (A) and abundance (B) in BMDM following IFN- γ stimulation (10 U/ml). Timecourse data are represented as described for Fig 3. * = Significant by MaSigPro test (see Materials and Methods and S1 Methods).

doi:10.1371/journal.pbio.1002364.g005

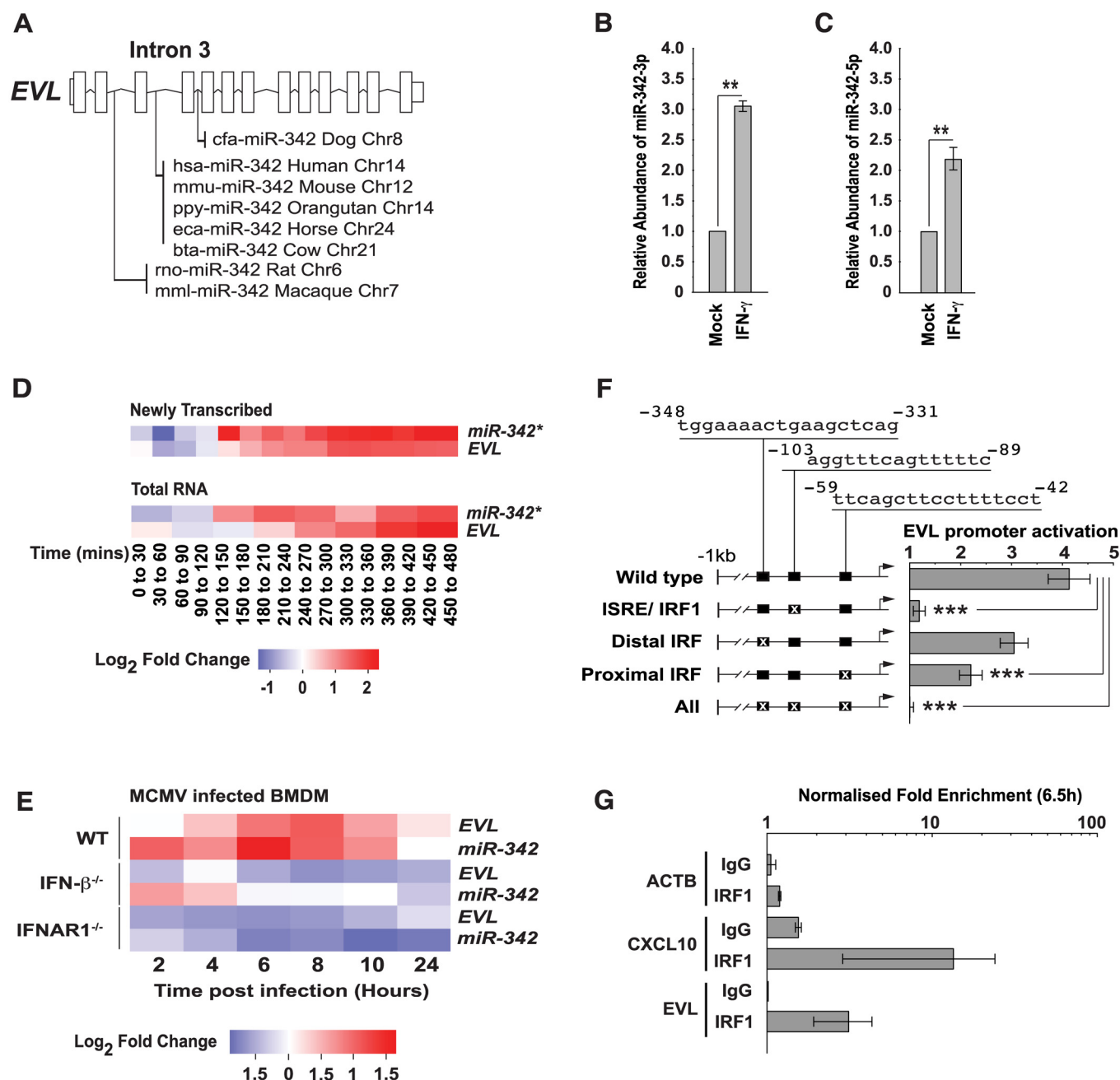


Fig 6. MiR-342 expression is directly coupled to IFN. (A) MiR-342 is encoded in the gene EVL and conserved between species. (B) Mature miRNA-342-3p expression relative to a small nucleolar RNA (SNORD47) in BMDM 24 h after treatment with 10 U/ml IFN- γ . Data are mean \pm SEM ($n = 3$). ** $p \leq 0.01$ by 1-sample t test. (C) Mature miRNA-342-5p expression relative to a small nucleolar RNA (SNORD47) in BMDM 24 h after treatment with 10 U/ml IFN- γ . Data are mean \pm SEM ($n = 10$, ** $p \leq 0.01$). (D) EVL and pri/pre-miR-342 RNA transcript synthesis and abundance in BMDM following IFN- γ stimulation (10 U/ml). Time course data are represented as described for Fig 1. * = Significant by MaSigPro test (see Materials and Methods and S1 Methods). (E) EVL and pri/pre-miR-342 (relative to $t = 0$) abundance in WT, IFNB1^{-/-} or interferon alpha receptor 1 (IFNAR1^{-/-}) BMDM infected with murine cytomegalovirus (MCMV) (multiplicity of infection [MOI] = 1). (F) Wild type or mutated 1 kb EVL promoter regions upstream of a luciferase reporter were used to test the relative importance of predicted ISRE, distal interferon regulatory factor (IRF), proximal IRF (or all) binding sites on IFN-induced expression ($n = 10$, *** $p \leq 0.001$). (G) Chromatin immunoprecipitation analysis of IRF1 transcription factor binding to *ACTB* (negative control), *CXCL10* (positive control), or the EVL promoter in BMDM cultured with IFN- γ (10 U/ml) for 6.5 h. Data are mean \pm SEM, $n = 2$.

doi:10.1371/journal.pbio.1002364.g006

containing a wild-type promoter or promoters with one or multiple mutations in the three predicted motifs validated the ISRE/IRF1 motif (Fig 6F). Mutation of either the proximal or distal interferon regulatory factor (IRF) motifs resulted in an intermediate reduction in promoter activity. We next performed chromatin immunoprecipitation assays to test for STAT1 (UniProt: P42225) and/or IRF1 (UniProt: P15314) binding to the murine *EVL*/miR-342 promoter. In IFN- γ -treated murine BMDM, the positive control CXCL10 promoter, but not the *EVL*/miR-342 promoter, was occupied by STAT1 or IRF1 2 h after treatment (S5B Fig upper graph). By 6.5 h, however, we observed an IRF1-specific enrichment (~3-fold) of *EVL*/miR-342 promoter sequences (Fig 6G) and by 24 h, IRF1 recruitment was still detectable (S5B Fig lower graph). We conclude that canonical IFN signalling is required for increased expression of miR-342 and that this involves the recruitment of IRF1 to the *EVL*/miR-342 promoter.

MiR-342-5p Regulates Sterol Biosynthesis

MiR-342 has recently been implicated in the regulation of SREBP2 in a cancer cell line; however, biological roles and precise mechanisms for the miRNA in relation to sterol biosynthesis and the immune response were not addressed [44]. To systematically test whether miR-155 or miR-342 could regulate the cholesterol pathway, miR-155, -342-3p or -342-5p mimics were transfected into a fibroblast cell line and RNA levels for selected cholesterol biosynthesis enzymes quantitated. While miR-155 and miR-342-3p had no significant effect on gene expression in the cholesterol pathway, miR-342-5p reduced the overall abundance of *HMGCS1* (–32%), *HMGCR* (–51%), *MVD* (–48%), *SQLE* (–51%) and *NSDHL* (Entrez Gene: 18194) (–62%) relative to cells treated with the nontargeting control (Fig 7A). In a comprehensive sterol pathway analysis of RNA extracted from primary murine embryo fibroblasts (pMEF) transfected with miR-342-3p or -5p, we proceeded to show that miR-342-5p but not miR-342-3p induced a significant, coordinate reduction in the abundance of 12 out of 19 sterol biosynthesis pathway transcripts analysed (Fig 7B and 7C). S6 Fig shows a comparison of the reduction in abundance of the sterol pathway transcripts by miR-342-5p with that elicited by a *SREBF2* targeting siRNA. Notably, the abundance of the fatty acid-associated transcript *FASN* (Entrez Gene: 14104) was not altered by miR-342-5p in these experiments (Fig 7B and 7C).

The concordant reduction of so many genes was consistent with the repression of a master regulator of the sterol biosynthesis pathway. SREBP2, the master transcriptional regulator for the majority of genes encoding sterol pathway enzymes, contains a predicted miR-342-5p binding site that is conserved in both human and mouse (Fig 8A). To directly test the function of this predicted miR-342-5p target, we generated dual-luciferase reporter constructs for the human and mouse *SREBF2* and mouse *SREBF1* 3'UTRs. Cotransfection of these reporters with miR-342-5p significantly reduced luciferase activity for the murine *SREBF2* but not the *SREBF1* reporter construct (Fig 8B, 8C and 8D). In addition, mutation of the seed region in both the human and mouse *SREBF2* 3'UTR increased luciferase expression (Fig 8B and 8C). Transfection of several mouse or human cell types with the miR-342-5p mimic decreased *SREBF2* transcript abundance (Fig 9A: pMEF, Fig 9B: BMDM, Fig 9D: NIH/3T3, S7A and S7B Fig). To avoid any complications arising from miRNA overexpression, we also sought to test the effect of inhibiting endogenous miR-342-5p on IFNG-mediated *SREBF2* regulation. In these experiments, we found miR-342-5p inhibitor significantly decreased but did not eliminate the repressive effects of a nonsaturating dose of IFNG [45] on *SREBF2* copy number in BMDM (Fig 9C). The results of these investigations show a contributory role of approximately 30% to 50% for miR-342-5p in the macrophage IFN suppression of *SREBF2*.

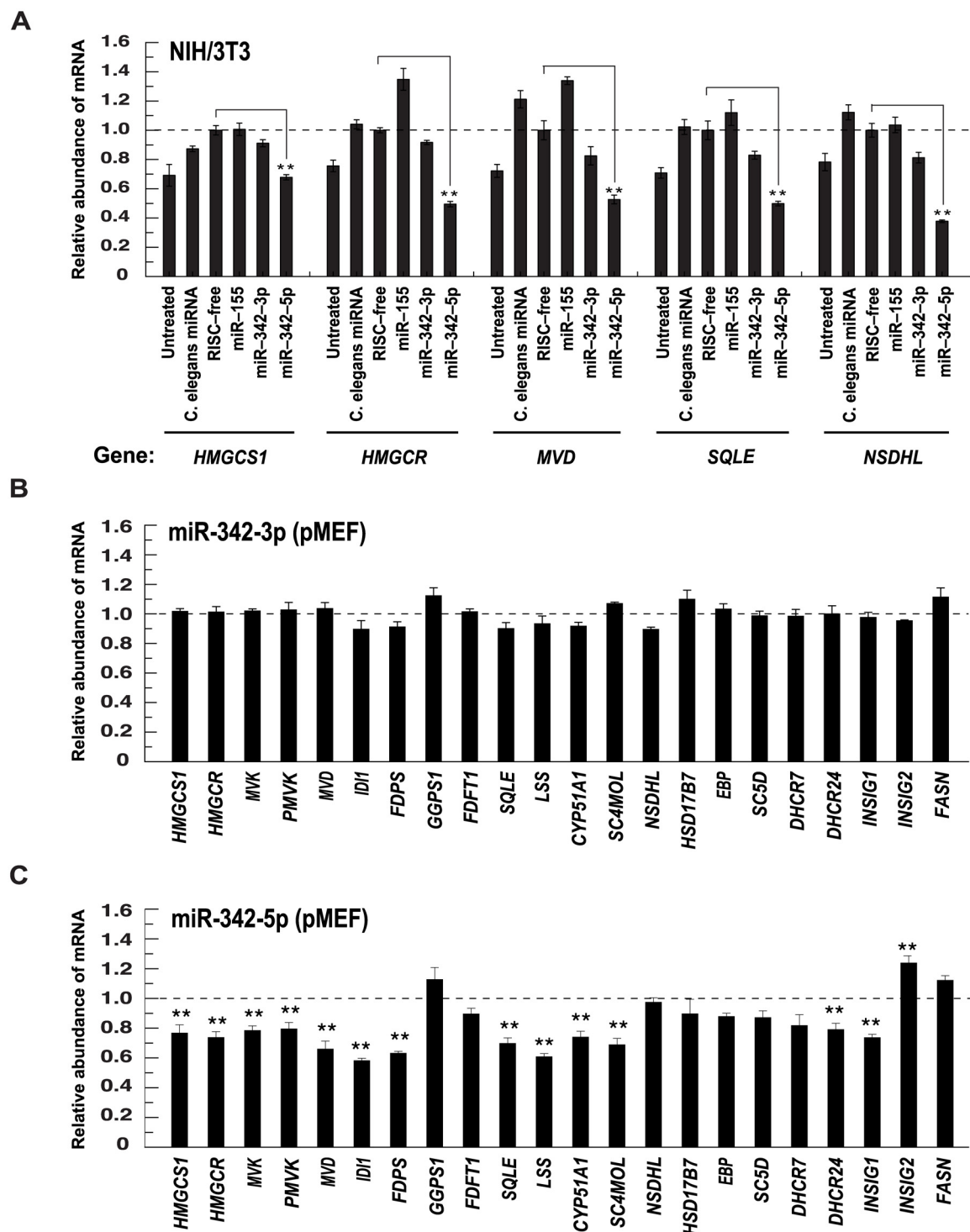


Fig 7. MiR-342-5p regulates the sterol pathway. (A) IFN-regulated miRNA effects on cholesterol pathway transcript abundance in NIH/3T3 fibroblasts. Data normalised to nontargeting siRNA and are mean \pm SEM ($n = 3$). ** $p \leq 0.01$. (B) MiR-342-3p effects on cholesterol pathway transcript abundance in pMEF. Data are normalised to nontargeting siRNA and are mean \pm SEM ($n = 3$). (C) MiR-342-5p effects on cholesterol pathway transcript abundance in pMEF. Data are normalised to nontargeting siRNA and are mean \pm SEM ($n = 6$), ** $p \leq 0.01$.

doi:10.1371/journal.pbio.1002364.g007

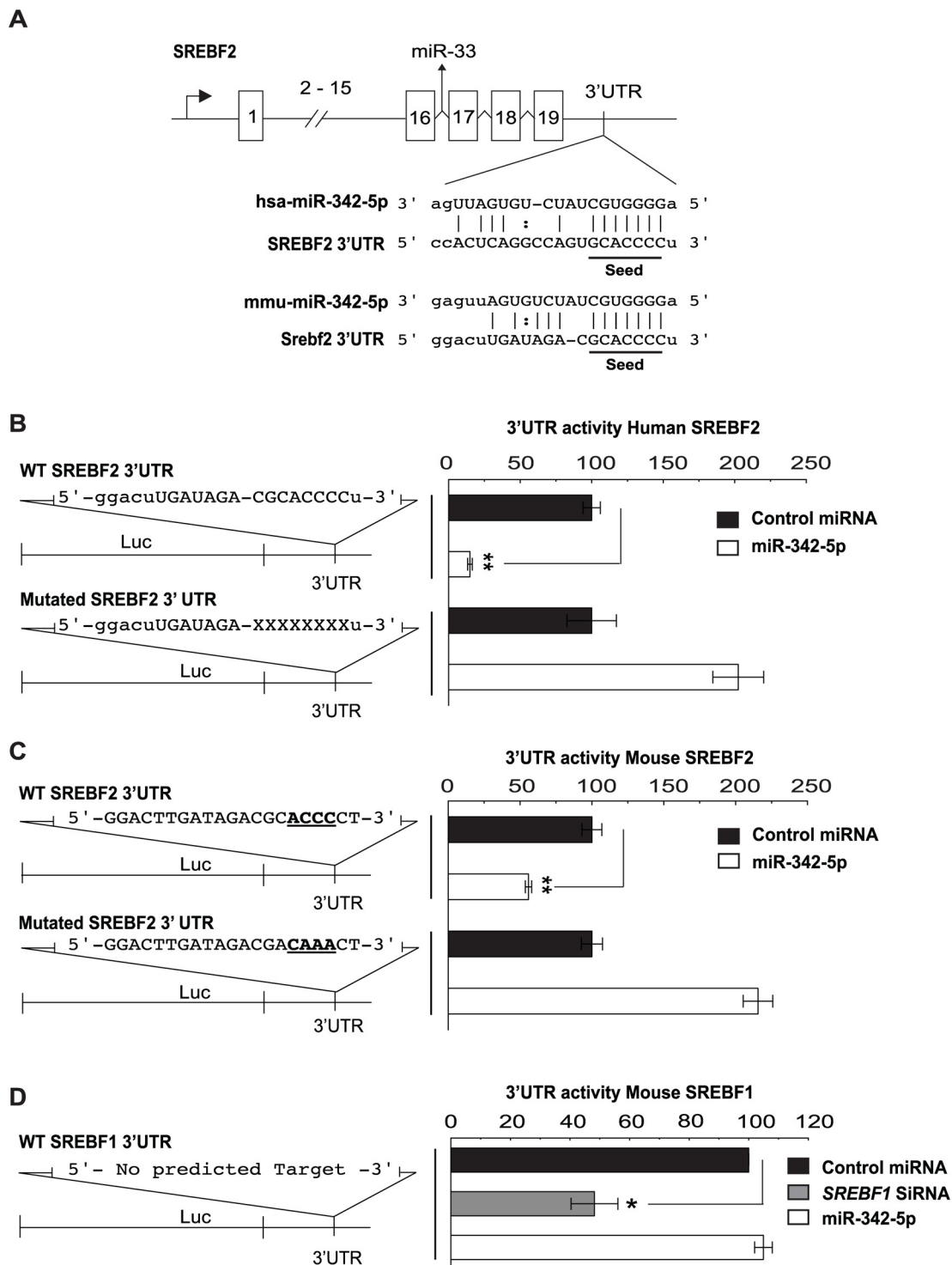


Fig 8. MiR-342-5p specifically targets the *SREBF2* transcript. (A) Predicted targets for miR-342-5p in human and mouse *SREBF2* transcripts. (B) Specific targeting of Human *SREBF2* 3'UTR by miR-342-5p. Data are mean \pm SEM ($n = 3$). $** p \leq 0.01$. (C) Specific targeting of Mouse *SREBF2* 3'UTR by miR-342-5p. Data are mean \pm SEM ($n = 3$). $** p \leq 0.01$. (D) MiR-342-5p does not target wild-type 3'UTR of murine *SREBF1*. Data are mean \pm SEM ($n = 3$). $* p \leq 0.05$.

doi:10.1371/journal.pbio.1002364.g008

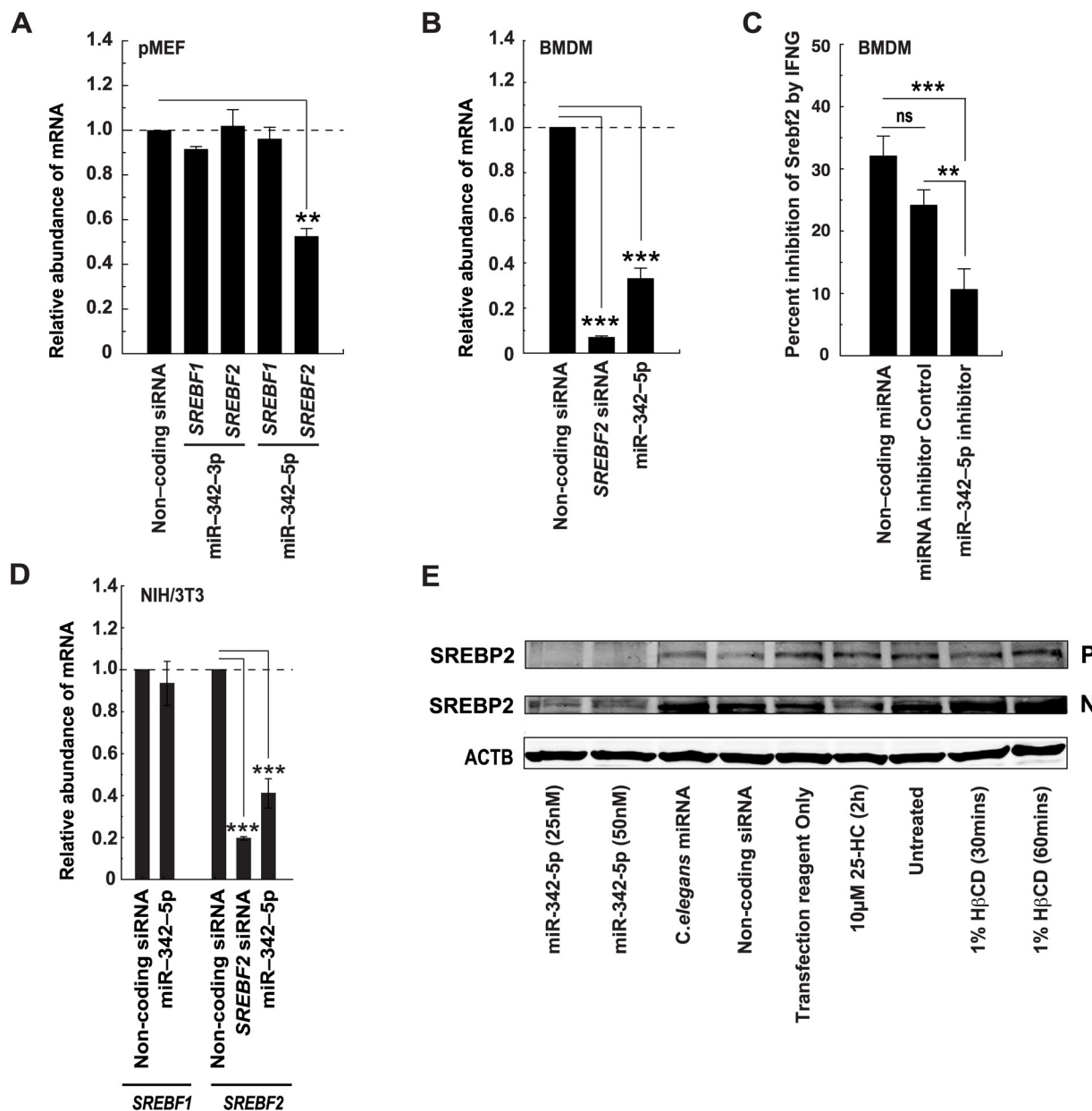


Fig 9. MiR-342-5p regulates SREBF2/SREBP2. (A) MiR-342-3p and -5p effects on *SREBF1* and *SREBF2* transcript abundance in pMEF. Data are mean \pm SEM (miR-342-3p $n = 3$, miR-342-5p $n = 6$), $** p \leq 0.01$). (B) MiR-342-5p effects on *SREBF2* transcript abundance in BMDM. Data are mean \pm SEM (*SREBF2* siRNA $n = 5$, miR-342-5p $n = 6$). $*** p \leq 0.001$. (C) A miR-342-5p inhibitor reduces the effects of IFNG-elicited *SREBF2* repression in BMDM. IFNG treatment was 2.5 ng/ml for 24 h; data are mean \pm SEM ($n = 6$ except *SREBF2* siRNA $n = 5$). $** p \leq 0.01$, $*** p \leq 0.001$. (D) MiR-342-5p effects on *SREBF1* and *SREBF2* transcript abundance in NIH/3T3 fibroblasts. Data normalised to nontargeting siRNA and are mean \pm SEM (*SREBF2* siRNA $n = 6$, miR-342-5p $n = 9$). $** p \leq 0.001$ by 1-sample t test. (E) Western blot analysis of Uncleaved (P) or cleaved (N) SREBP2 protein changes due to miR-342-5p, *Caenorhabditis elegans* miRNA, RISC-free siRNA, Transfection Reagent Only, 10 μ M 25-HC for 2 h, 1% H β CD for 30 min or 1% H β CD for 60 min in NIH/3T3 cells.

doi:10.1371/journal.pbio.1002364.g009

In further experiments, protein levels for both the uncleaved (P) and cleaved (N) form of the SREBP2 (UniProt: Q3U1N2) protein were also decreased in cells transfected with the miR-342-5p mimic (Fig 9E). The latter result contrasts with the action of 25-HC that reduced the cleaved protein (N) abundance alone (Fig 9E). Human *SREBF1* (Entrez Gene: 6720) has

recently been identified as a target of miR-342-5p (confirmed in [S7B Fig](#)) [44]. Our bioinformatics analysis did not identify a target for this miRNA anywhere in the murine transcript, and the transfection of mouse fibroblasts with a miR-342-5p mimic did not elicit a significant decrease in overall SREBF1 transcript abundance ([Fig 9A and 9D](#)).

The transcript *SREBF2* contains an intronic miRNA (miR-33) that regulates fatty acid degradation and cholesterol homeostasis [32]. In this connection, it is worth noting that type 1 and 2 IFN treatment and transfection of miR-342-5p reduced miR-33 abundance in BMDM ([Fig 10A](#) and [S7C Fig](#)). Since miR-342-5p targets the mature, spliced *SREBF2* transcript in the cytoplasm, the reduction of miR-33 must be occurring in the nucleus prior to splicing. Whilst the underlying molecular mechanism for this remains to be fully elucidated, we anticipate a decrease in miR-33 occurs as a result of miR-342-5p effects on the transcriptional autoregulation of the *SREBF2* promoter by SREBP2 ([Fig 10D](#)). Since miR-33-5p targets *ABCA1* (Entrez Gene: 11303) and *ABCG1* (Entrez Gene: 11307) in mice [32], the question arose: can miR-342-5p regulate the abundance of these cholesterol efflux-related transcripts? The nuclear hormone Liver X receptors LXR α (NR1H3 UniProt: Q9Z0Y9) and LXR β (NR1H3 UniProt: Q60644) are activated by oxysterol binding and are well-known regulators of cholesterol homeostasis acting to regulate the transcription of *ABCA1* and *ABCG1*. As a consequence, they mediate cholesterol efflux from the cell [46]. To analyse miR-342-5p modulation of *ABCA1* and *ABCG1* expression, we measured the abundance of these transcripts in transfected BMDM treated with the LXR agonist T0901312. A schematic illustrating the logic of this approach is presented ([S7D Fig](#)). MiR-342-5p suppression of miR-33 was confirmed by Q-RT-PCR in these experiments ([S7E Fig](#)) and, as previously reported, miR-33 significantly reduced the effects of LXR activation on *ABCA1* and *ABCG1* transcript abundance in our experiments ([Fig 10B and 10C](#)) [32]. In contrast, miR-342-5p enhanced the effect of T0901312 treatment on *ABCA1* and, significantly, on *ABCG1* expression ([Fig 10B and 10C](#)).

We conclude that miR-342-5p targets and acts to suppress *SREBF2* via a single, conserved miRNA binding site in its 3'UTR. In addition, our results point to a coupling of miR-342-5p to miR-33 regulation—likely through the SREBP2 transcriptional autoregulation pathway [29]. MiR-342-5p, therefore, also has the potential to counter-regulate LXR-mediated control of cholesterol homeostasis.

We next investigated whether miR-342-5p could regulate intracellular sterol levels. Mass spectrometry analyses of transfected BMDM showed miR-342-5p-induced reductions in metabolites from the mevalonate-shunt (24S,25-epoxycholesterol, [Fig 11A](#)), Bloch (desmosterol, [Fig 11B](#)) and Kandutsch-Russell pathways (7-dehydrocholesterol + 8(9)-dehydrocholesterol, [Fig 11C](#)) demonstrating miR-342-5p inhibits sterol biosynthesis. Accordingly, free ([Fig 11D](#)) and total cholesterol ([S8A Fig](#)) were also reduced by miR-342-5p transfection in BMDM and fibroblasts, respectively. In agreement with our gene expression data, we further showed that an inhibitor of endogenous miR-342-5p could moderately increase total cholesterol in transfected fibroblasts ([S8B Fig](#)). In these experiments, miR-342-5p consistently reduced intracellular cholesterol concentration in cells cultured in medium containing 10% serum. This agrees with experiments showing that miR-342-5p targets biosynthesis through *SREBF2* and probably also influences other components of the cholesterol regulatory network responsible for influx and/or efflux, e.g., *ABCG1*.

Our previous studies have indicated that although IFN suppression of sterol biosynthesis and the antiviral activities of 25-HC are partly SREBP-dependent, SREBP independent sterol pathway-related mechanisms predominate in the repression of virus infection [18]. In this study, candidate targets for miR-342-5p in the sterol pathway members *ID11* (Entrez Gene: 319554), *SC4MOL* (MSMO1 Entrez Gene: 66234), and *DHCR7* (Entrez Gene: 13360) suggested a possible SREBP-independent mechanism for miR-342-5p ([Fig 12A](#) and [S1 Table](#)). *ID11*

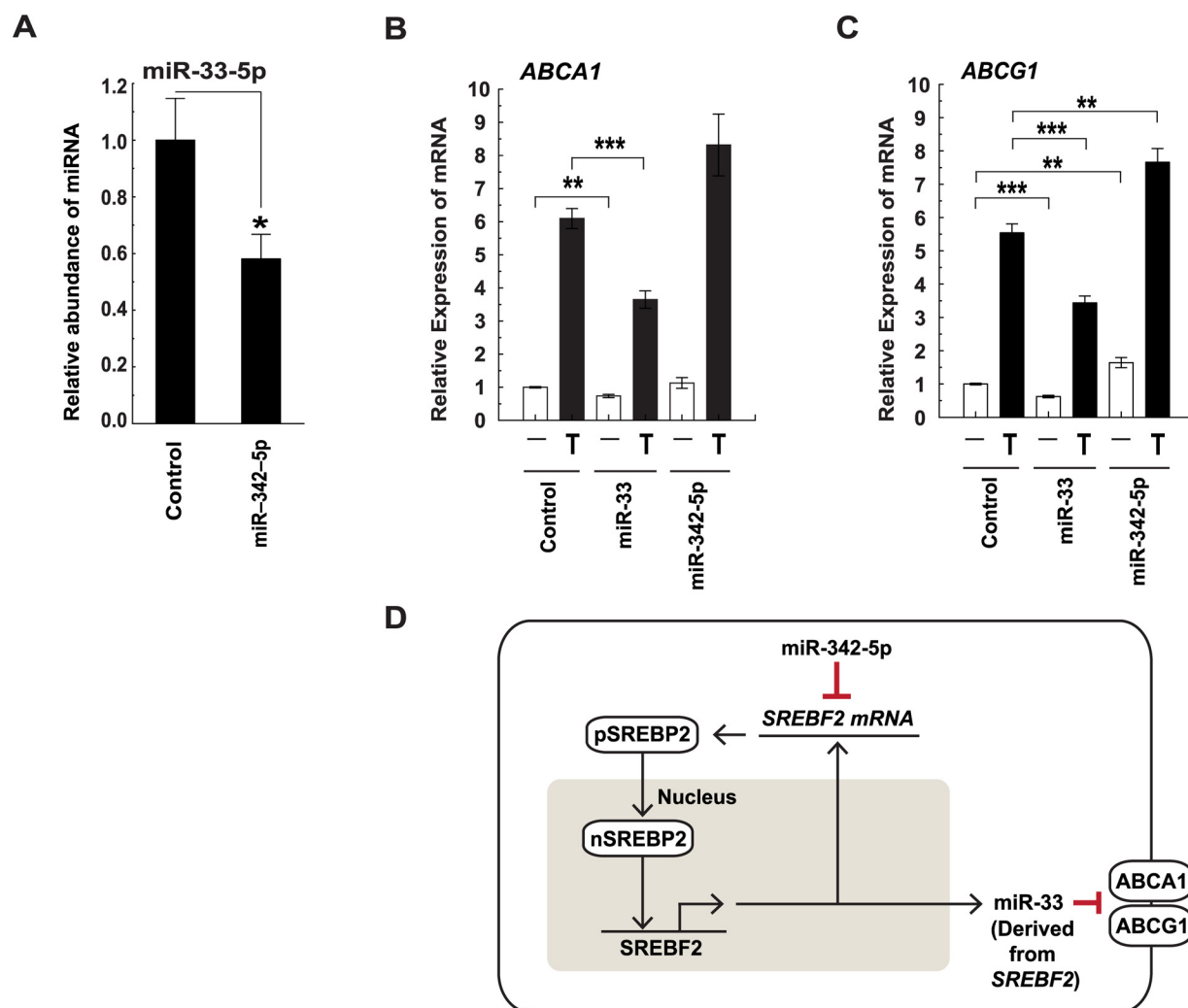


Fig 10. MiR-342-5p regulates miR-33 expression and function. (A) MiR-342-5p effects on miR-33-5p abundance. Data are mean \pm SEM ($n = 3$) * $p \leq 0.05$ by 1-sample t test. (B) MiR-342-5p effects on *ABCA1* transcript abundance in vehicle (-) or T0901317 (T) treated BMDM. Data are mean \pm SEM ($n = 6$). ** $p \leq 0.01$, *** $p \leq 0.001$ in comparisons between identical LXR treatments in control versus miRNA-treated cells. (C) MiR-342-5p effects on *ABCG1* transcript abundance in vehicle (-) or T0901317 (T)-treated BMDM. Data are mean \pm SEM ($n = 6$). ** $p \leq 0.01$, *** $p \leq 0.001$ in comparisons between identical LXR treatments in control versus miRNA-treated cells. (D) Schematic showing proposed mechanism of miR-33 regulation and its downstream effects on *ABCA1* and *ABCG1*, via miR-342-5p modulation of the SREBP2 autoregulatory loop.

doi:10.1371/journal.pbio.1002364.g010

(UniProt: P58044) in particular plays a key role in the synthesis of isoprenoids from sterol pathway metabolic intermediates—a process of great significance to a range of viruses [18,47]. Dual luciferase assays were, therefore, used to validate miRNA target sites for miR-342-5p in the 3'UTR of *ID11* and *SC4MOL* (Fig 12B and 12C). MiR-342-5p also reduced the expression of a WT *DHCR7* 3' UTR reporter (S8C Fig). Taken together, our data show that the IFN-induced miR-342-5p targets sterol metabolism at multiple levels in both a SREBP2-dependent and -independent manner and can regulate both cholesterol biosynthesis and homeostasis.

MiR-342-5p Has Broad-Spectrum Antiviral Activity In Vitro and In Vivo

We, and others, have previously demonstrated that sterol pathway regulation is an integral part of the IFN antiviral response [18,24,46]. We speculated; therefore, that the regulation of

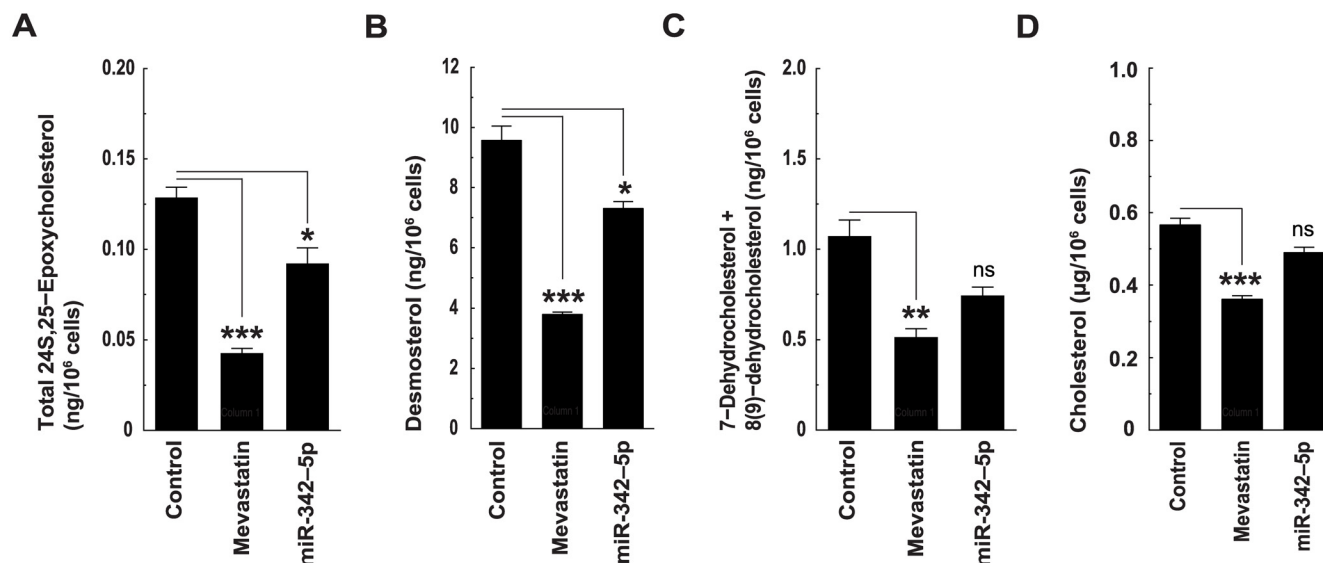


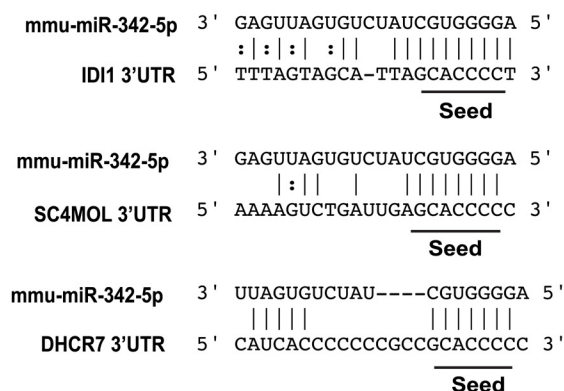
Fig 11. MiR-342-5p regulates intracellular cholesterol homeostasis. (A to D) Mass spectrometry quantitation of total 24S,25-Epoxycholesterol (A), Desmosterol (B), 7-Dehydrocholesterol + 8(9)-dehydrocholesterol (C) and free cholesterol (D) in BMDM transfected with miR-342-5p for 48 h. Data are mean \pm SEM ($n = 3$). * $p \leq 0.05$, ** $p \leq 0.01$, *** $p \leq 0.001$.

doi:10.1371/journal.pbio.1002364.g011

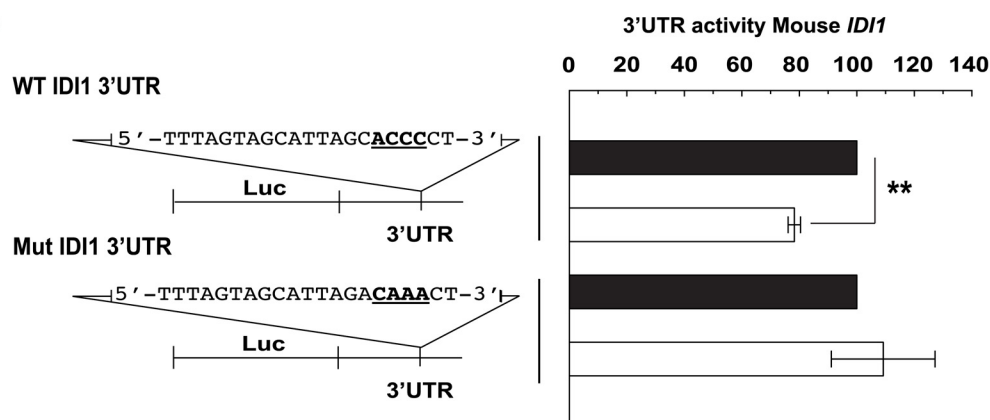
sterol biosynthesis and homeostasis by miR-342-5p might mediate antiviral functions and protect cells from virus infection. In support of this hypothesis, we found miR-342-5p but not miR-342-3p significantly inhibited growth of MCMV (S9A Fig), and that this occurred in a dose-dependent manner (Fig 13A). We further showed that an inhibitor of endogenous miR-342-5p could partially rescue MCMV replication in NIH/3T3 fibroblasts treated with IFN- γ (Fig 13B, IFN- γ function confirmed in S9B Fig). We next tested whether miR-342-5p is capable of inhibiting MCMV in vivo. BALB/c mice were injected intraperitoneally with 10 μ g (total) of miR-342-5p mimic, inhibitor, or a nonspecific control RNA. Four d postinfection, MCMV titres in miR-342-5p-treated mice (relative to the nonspecific control miRNA) were significantly reduced ($\sim 1 \log_{10}$) in the kidney and lung (Fig 13C). In further, independent in vivo experiments, antiviral miR-342-5p effects were consistently observed; however, organs with a reduced virus titre varied between runs. This is likely attributable to variability in the delivery of relatively small doses of the miRNA (S9C and S9D Fig). In our in vivo experiments, the miR-342-5p inhibitor did not increase viral replication relative to the negative control miRNA treatment (Fig 13C). While further genetic loss-of-function studies are required to unequivocally investigate the in vivo function of miR-342-5p, these studies support the view that this miRNA imparts antiviral activity in vivo and in vitro.

We have previously reported that IFN-induced effects arising via sterol pathway regulation play a key role in limiting MCMV replication and are mediated via the isoprenoid-prenylation branch [18]. This results in severely restricted viral spread at a postentry stage of infection. In agreement with our 25-HC-related work, miR-342-5p transfection of cells did not alter viral entry (Fig 14A) but did significantly reduce infectious virus production (Fig 14B), immediate early gene expression (Fig 14C and 14D), and MCMV plaque diameter. Fig 14E shows a miR-342-5p elicited reduction in the diameter of plaques at 3 d post infection, whilst Fig 14F shows data from a quantitative analysis of plaque diameter in both WT and STAT1^{-/-} cells at 4dpi. The latter cell type was used to minimise potential side effects of transfection on immune activation and subsequent antiviral responses. Of note, miR-342-5p did not notably alter the viability of cells used in any of the described analyses (S10A and S10B Fig). Since miR-342-5p

A



B



C

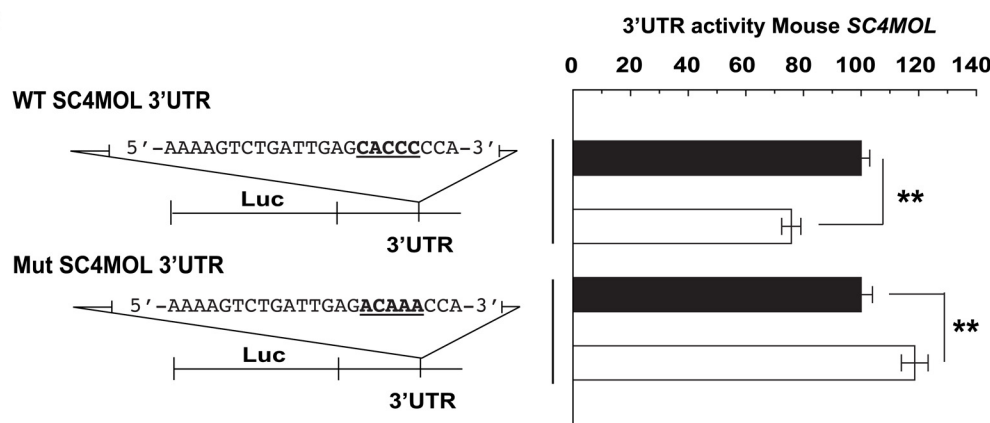


Fig 12. Multihit targeting of the sterol biosynthesis pathway by miR-342-5p. (A) Predicted targets for miR-342-5p in mouse *IDI1*, *SC4MOL*, and *DHCR7* transcripts. (B) Specific targeting of Mouse *IDI1* 3'UTR by miR-342-5p. Data are mean \pm SEM ($n = 3$), ** $p \leq 0.01$. (C) Specific targeting of Mouse *SC4MOL* 3'UTR by miR-342-5p. Data are mean \pm SEM ($n = 3$), ** $p \leq 0.01$.

doi:10.1371/journal.pbio.1002364.g012

targets the sterol metabolic network, we tested whether it exerted broader antiviral activity by analysing its effects on Human Cytomegalovirus (HCMV), Herpes Simplex virus 1 (HSV1), and Influenza A virus (H1N1) (Fig 15A, 15B and 15C). Replication of all three viruses was inhibited by miR-342-5p (80% for HSV-1, 50% for HCMV and 60% for Influenza virus A).

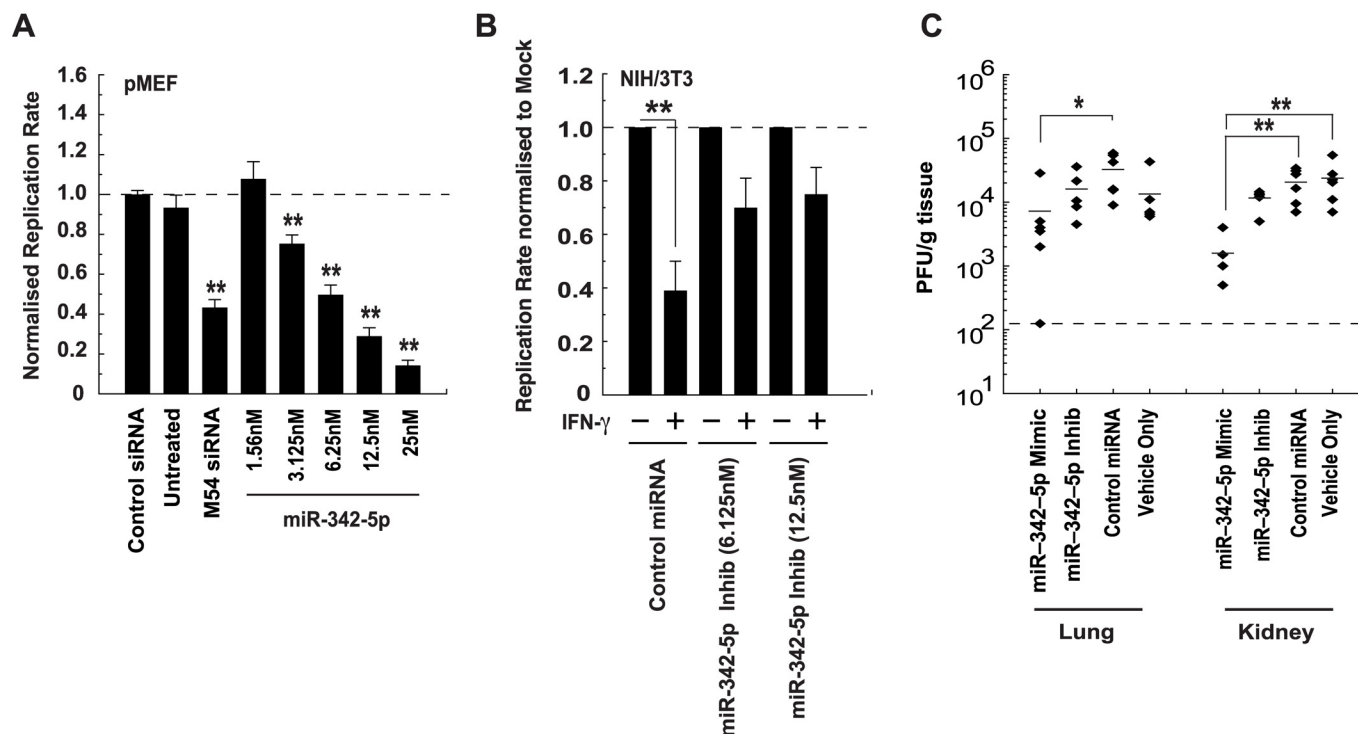


Fig 13. MiR-342-5p is antiviral. (A) MCMV-GFP replication is inhibited in pMEF by miR-342-5p in a dose-dependent manner. Data are normalised to values obtained with nontargeting control siRNA and are mean \pm SEM of slope between 62 and 92 hpi ($n = 6$), ** $p < 0.01$. (B) An inhibitor of miR-342-5p reduces the antiviral effect of IFN- γ . NIH/3T3 cells were transfected with 6.125 or 12.5 nM miR-342-5p inhibitor for 48 h. Cells were then mock or IFN- γ treated (100 pg/ml) for 6 h, infected with MCMV-GFP (MOI = 0.0025), and then cultured in medium containing 3% delipidised serum. Data show replication slope of virus in IFN- γ treated cells relative to Mock. Data are mean \pm SEM of slope between 62 and 92 hpi (control miRNA $n = 4$, inhibitor $n = 6$). ** $p \leq 0.01$. (C) MCMV titre (4 dpi) in lung and kidney of BALB/c mice treated with the miR-342-5p, miR-342-5p ZEN-AMO inhibitor or control miRNA. Data are titre in organ of individual mouse ($n = 6$). * $p \leq 0.05$, ** $p \leq 0.01$.

doi:10.1371/journal.pbio.1002364.g013

Broad Differential Antiviral Effects of miR-342-5p Are Mediated through Multihit Targeting of the Sterol Pathway

As previously discussed, whilst IFN-induced suppression of sterol biosynthesis involves a SREBP-dependent mode of action, alternative sterol pathway-related mechanisms predominate in the repression of virus replication [18]. With this in mind, we sought to further investigate SREBP-independent antiviral mechanisms for miR-342-5p. For these experiments, we evaluated whether the specific knockdown of predicted non-SREBP targets of miR-342-5p (*ID11*, *DHCR7* and *SC4MOL*) alone were sufficient to impart antiviral effects observed for the miRNA (Fig 16). Knockdown of specific targets by these siRNA was confirmed in pMEF by Q-RT-PCR (S9F Fig). Notably, siRNA-mediated knockdown of *ID11* consistently inhibited (~70%) MCMV replication in pMEF with a similar magnitude as inhibition mediated by a virus-specific (M54, MCMV DNA polymerase) or *HMGCR*-targeting siRNA (Fig 16). In contrast, siRNA targeting of *SC4MOL* and *DHCR7*—enzymes that function downstream of the isoprenoid pathway branch—did not limit MCMV replication. Fig 12C shows functional targeting of *SC4MOL*. Since *SC4MOL* catalyses six distinct enzymatic steps in the Kandutsch-Russell/Bloch arms of the sterol biosynthesis pathway (Fig 4), we anticipate miR-342-5p targeting of this enzyme could have significant effects on cholesterol biosynthesis in the cell and antiviral effects on pathogens dependent on this arm of the pathway. This data is in agreement

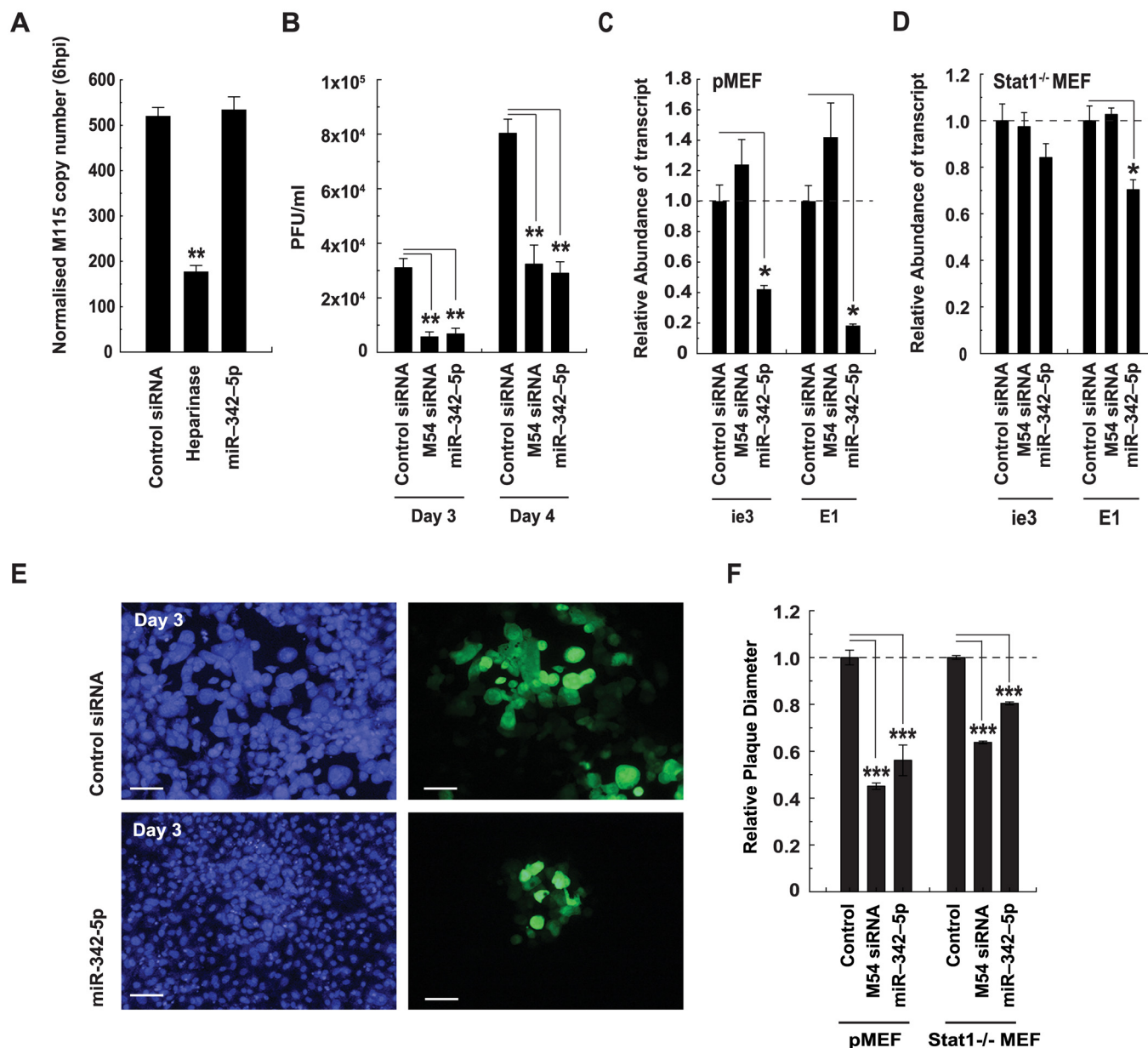


Fig 14. MiR-342-5p inhibits infectious virus production and spread. (A) MiR-342-5p does not affect MCMV entry. MCMV M115 DNA copy number in pMEF transfected with negative control siRNA or miR-342-5p (25 nM) for 48 h and then infected with MCMV (MOI = 0.1) for 4 h. Data are mean \pm SEM ($n = 3$). ** $p \leq 0.01$. (B) MiR-342-5p inhibits infectious MCMV production. PMEF were transfected (25 nM, 48 h) with nontargeting control siRNA, MCMV M54 siRNA, or miR-342-5p and then infected with MCMV (MOI = 0.0025). PFU in supernatants were then quantitated by plaque assay at 3 dpi and 4 dpi. Data are mean \pm SEM ($n = 3$). ** $p \leq 0.01$. (C and D) MiR-342-5p inhibits MCMV gene expression. Primary MEF (C) or STAT1^{-/-} MEF (D) were transfected (25 nM, 48 h) with nontargeting control siRNA, MCMV M54 siRNA, or miR-342-5p and then infected with MCMV (MOI = 0.01). MCMV transcript abundance was then quantitated by Q-RT-PCR at 6 hpi. Data are mean \pm SEM (WT, $n = 3$, STAT1^{-/-} $n = 4$). * $p \leq 0.05$. (E) Images showing effects of miR-342-5p on MCMV-GFP infected cells at 3 dpi. Bar = 100 μ m. (F) MiR-342-5p effects on plaque formation were analysed at 4 dpi when monolayers were fixed and plaque diameter quantitated using ImageJ software. Data are mean \pm SEM, $n = 4$. For pMEF: Control = 168 plaques counted, M54s siRNA 66 plaques counted, miR-342-5p $n = 23$. STAT1^{-/-} MEF: Control = 1,091 plaques counted, M54 siRNA = 822 plaques counted, miR-342-5p = 1,011 plaques counted. *** $p \leq 0.001$

doi:10.1371/journal.pbio.1002364.g014

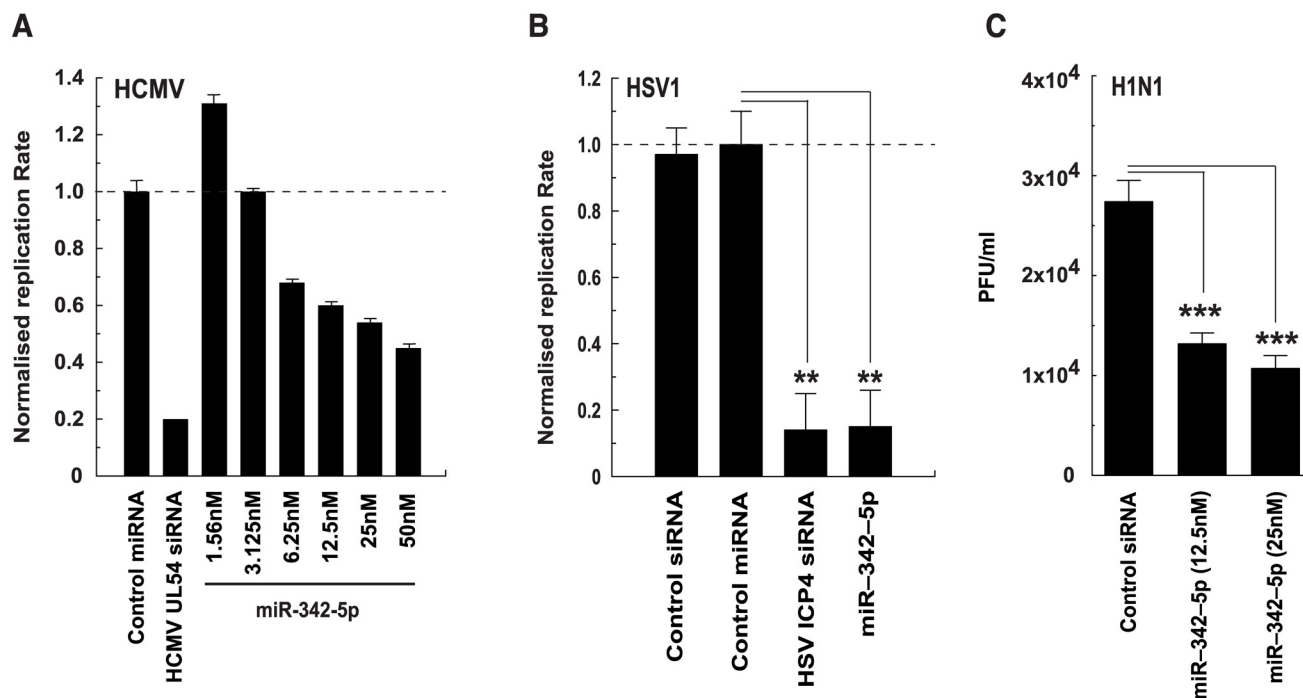


Fig 15. MiR-342-5p exerts a broad antiviral activity. (A) Inhibition of HCMV-GFP in MRC5 cells. Data are normalised to values obtained with *C. elegans* miRNA and are mean \pm SEM ($n = 2$). (B) Inhibition of HSV1-GFP in HeLa cells. Data are normalised to values obtained with *C. elegans* control miRNA and are mean \pm SEM ($n = 3$), ** $p \leq 0.01$. (C) Inhibition of Influenza virus (H1N1). A549 cells were transfected with miRNA for 48 h then infected with H1N1 for 24 h. Infectious virus titre in supernatants was then analysed by plaque assay. Data are mean \pm SEM. Control siRNA and miR-342-5p (25 nM) $n = 14$. MiR-342-5p (12.5 nM) $n = 6$. *** $p \leq 0.001$.

doi:10.1371/journal.pbio.1002364.g015

with previous studies and demonstrates that SREBP-independent targeting of the sterol biosynthesis pathway can elicit a profound antiviral effect [18,27].

It is known that unrelated viruses have a common dependency on the mevalonate-sterol pathway. Evidence suggests, however, that the relative significance of individual pathway members differs according to virus type and the respective demands of its replication strategy [48,49]. In the case of MCMV, we have previously demonstrated that the addition of exogenous metabolic intermediates (mevalonate or geranylgeranyl diphosphate but not squalene [SQL]) to cells can partially rescue MCMV growth in vitro by compensating for IFN or 25-HC-induced suppression of the sterol pathway [18,24]. In this study, we sought to test whether the antiviral effects of miR-342-5p require the suppression of metabolic intermediates in the mevalonate and sterol biosynthesis arms of the pathway. In a series of MCMV metabolite rescue experiments, and in agreement with our 25-HC-related work, we found that geranylgeraniol (GGOH) (Fig 17C and 17D) and, to a lesser degree, mevalanolactone (MEV) (Fig 17A and 17B) (but not farnesol [FOH] or SQL, Fig 17E and 17F), could significantly increase and partially restore MCMV growth in miR-342-5p transfected pMEF. These results highlight the importance of host-targeting of the mevalonate arm of the sterol pathway by miR-342-5p and are in good agreement with our previous studies [18,24].

In contrast to MCMV, a series of Influenza virus metabolite rescue experiments demonstrated that MEV, FOH, SQL and, to a lesser extent, GGOH all partially rescued infectious virus production by A549 cells in the context of miR-342-5p but not RAB11A siRNA transfection (Fig 18A, 18B, 18C and 18D). RAB11 (UniProt: P62491) is known to be required for Influenza A virus budding and filament formation [50]. Whilst caution should be taken to avoid

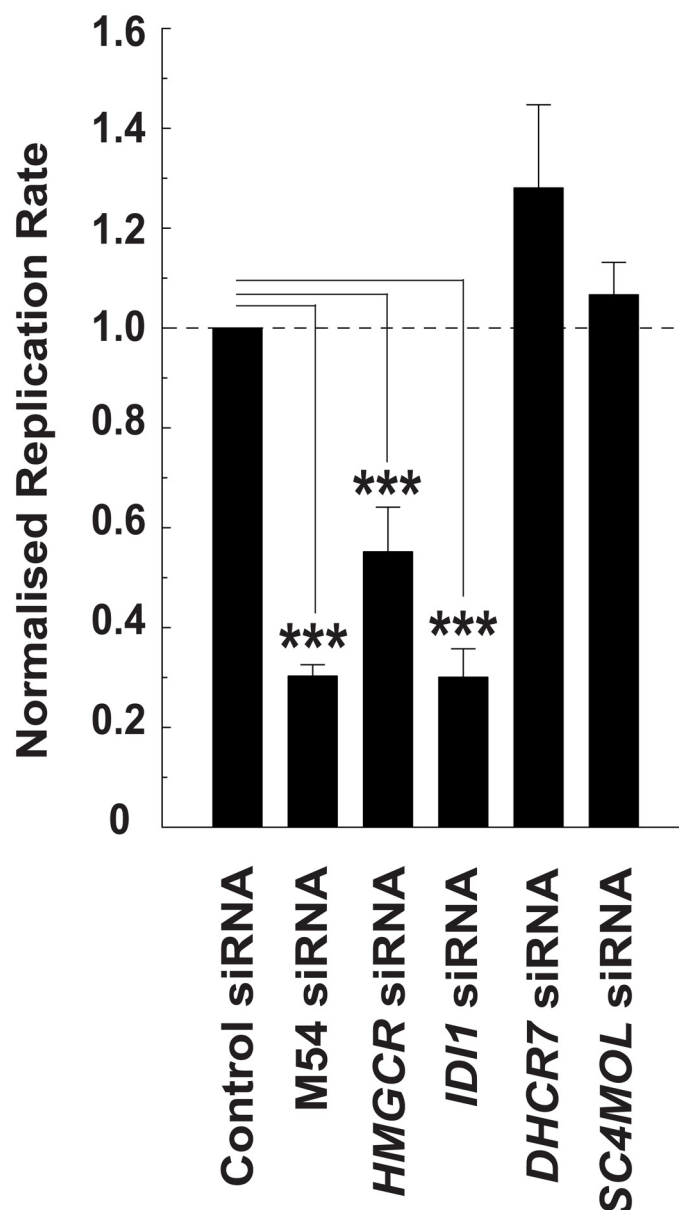


Fig 16. Knockdown of *IDI1* but not *DHCR7* or *SC4MOL* results in inhibition of MCMV replication in pMEF. MCMV-GFP replication in pMEF transfected with *HMGCR*, *IDI1*, *DHCR7* or *SC4MOL*-specific siRNA. Data are mean \pm SEM, ($n = 3$) *** $p \leq 0.001$.

doi:10.1371/journal.pbio.1002364.g016

overinterpreting the relative effects of specific metabolites in unrelated cell types, when taken together, our metabolite rescue data emphasise two key mechanistic issues. Firstly, unrelated viruses depend on distinct metabolic aspects of the sterol biosynthesis pathway for their replication. Secondly, despite these distinctions, “multihit” targeting of the sterol pathway by miR-342-5p enables this miRNA to elicit a broad antiviral effect.

MiR-342-5p has recently been reported to specifically target Cocksackie B virus [51]. While we cannot exclude the possibility that miR-342-5p also directly targets MCMV, HCMV, and/or HSV-1 (S2 Table), we failed to find any direct viral-RNA targets for the miRNA in Influenza virus A. Given the relatively low number of cellular miRNAs shown to have antiviral activity

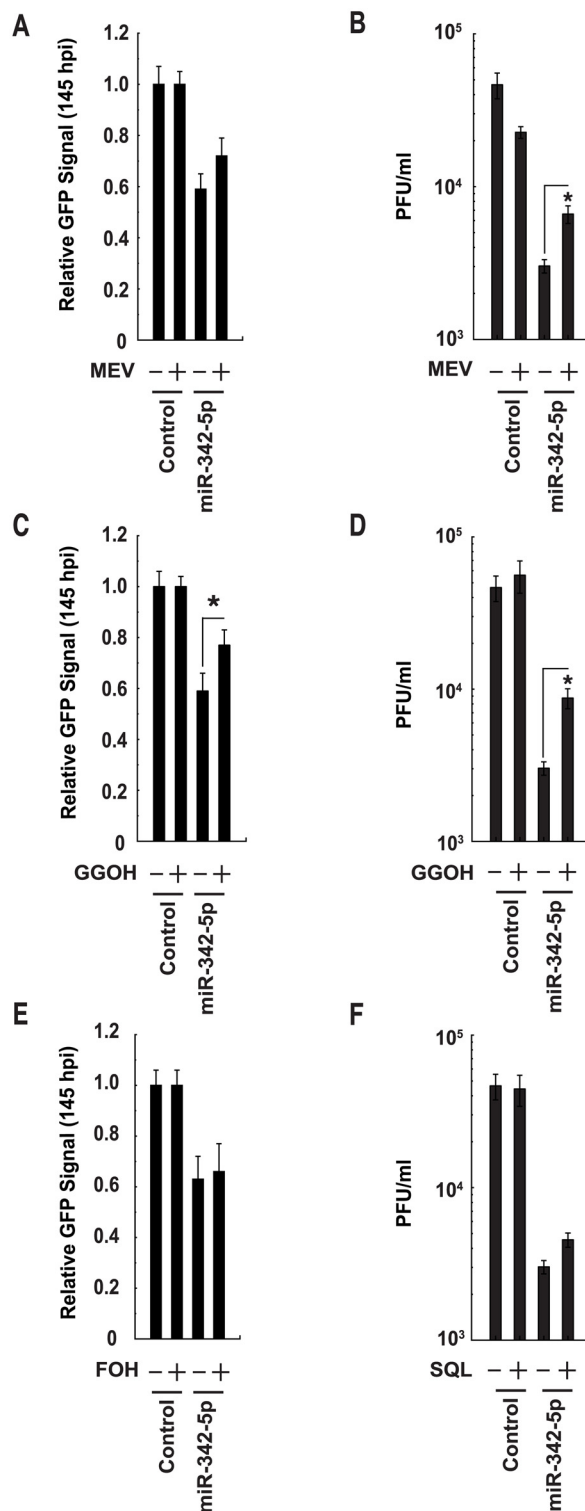


Fig 17. Inhibitory effects of miR-342-5p on MCMV are mediated through regulation of the mevalonate-isoprenoid pathway. (A) MCMV-GFP replication in pMEF transfected with control miRNA or miR-342-5p and then cultured with MEV (1 μ M final concentration). Data are mean \pm SEM ($n = 39$). (B) MCMV PFU production in pMEF transfected with control miRNA or miR-342-5p and then cultured with MEV (20 μ M final concentration). Data are mean \pm SEM ($n = 4$). * $p \leq 0.05$. (C) MCMV-GFP replication in pMEF transfected with control miRNA or miR-342-5p and then cultured with GGOH (1 μ M final concentration). Data are mean

+/- SEM ($n = 39$). * $p \leq 0.05$. (D) MCMV PFU production in pMEF transfected with control miRNA or miR-342-5p and then cultured with GGOH (20 μ M final concentration). Data are mean +/- SEM ($n = 4$). * $p \leq 0.05$. (E) MCMV-GFP replication in pMEF transfected with control miRNA or miR-342-5p and then cultured with FOH (1 μ M final concentration). Data are mean +/- SEM ($n = 19$). (F) MCMV PFU production in pMEF transfected with control miRNA or miR-342-5p and then cultured with SQL (20 μ M final concentration). Data are mean +/- SEM ($n = 3$). * $p \leq 0.05$.

doi:10.1371/journal.pbio.1002364.g017

[5], we consider it unlikely that miR-342-5p exerts broad antiviral effects by directly targeting viral transcripts. On the basis of these observations, combined with evidence from the metabolic rescue experiments, we propose that miR-342-5p exerts its broad antiviral effects by targeting host metabolic activities associated with the mevalonate-sterol biosynthesis pathway.

Discussion

In this study, we identify miR-342-5p as a cellular miRNA with broad antiviral properties whose transcriptional regulation is coupled to IFN signalling by IRF1. MiR-342-5p is, therefore, an integral component of the cell-autonomous IFN response and exerts its antiviral effects by inhibiting the sterol metabolic network through SREBP-dependent and -independent mechanisms. The latter are, at least in part, mediated via the targeting of key genes in the sterol metabolic network. Metabolic rescue experiments show that unrelated viruses have a common requirement for the mevalonate biosynthesis arm yet depend on subtly different aspects of the distal sterol pathway for their life cycle.

In its role, miR-342-5p complements and reinforces the antiviral functions of the rapidly induced oxysterol 25-HC on sterol biosynthesis—in particular affecting the mevalonate-isoprenoid branch of the pathway. [S11 Fig](#) summarises our current understanding of the

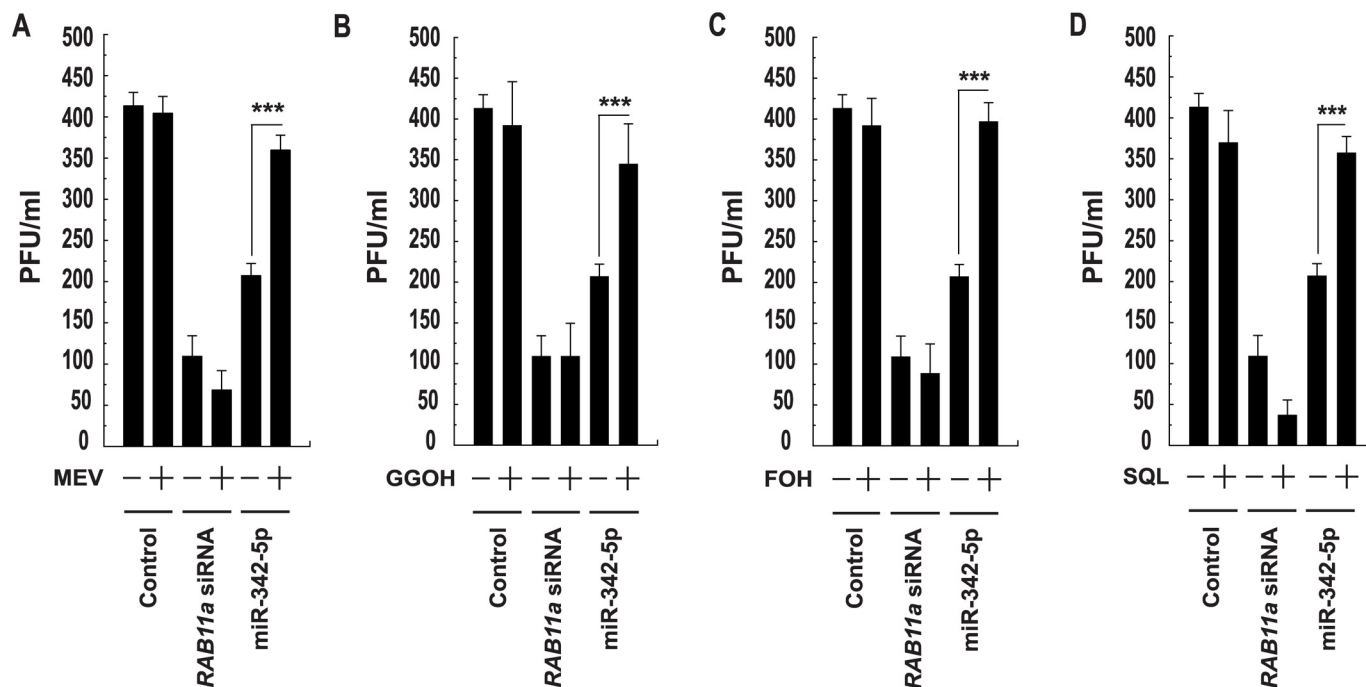


Fig 18. Effects of miR-342-5p on Influenza virus are mediated through regulation of the mevalonate and sterol arms of the biosynthesis pathway. (A to D) Influenza virus A (H1N1) PFU production in A549 cells transfected with control miRNA or miR-342-5p for 48 h and then infected with H1N1 (MOI = 0.1) and cultured with MEV (A, 10 μ M, $n = 8$), GGOH (B, 10 μ M, $n = 12$), FOH (C, 10 μ M, $n = 12$) or SQL (D, 10 μ M, $n = 12$) for 24 h.

doi:10.1371/journal.pbio.1002364.g018

mechanisms by which IFN induced 25-HC and miR-342-5p function to coordinately regulate sterol biosynthesis. In murine BMDM, synthesis of *CH25H* mRNA is controlled by STAT1 and is up-regulated in the first 30 min after cells are activated by IFN- γ [18]. In contrast, *EVL* and pri/pre-miR-342 RNA expression increases 2 h to 3 h after IFN- γ activation of BMDM and is regulated by IRF1. Our data demonstrate, therefore, an IFN-elicited sequential regulation of the sterol metabolic network in which 25-HC provides an immediate, rapid mechanism for decreasing sterol biosynthesis and mediating antiviral effects. This involves a 25-HC blockade of SREBP2 translocation to the nucleus and the proteolytic degradation of HMGCR [18,52]. The effects of 25-HC are followed by miR-342-5p further promoting a more sustained fine-tuning of sterol metabolism and antiviral effects in the cell by targeting *SREBF2* RNA and transcripts encoding select enzymes of the sterol biosynthesis pathway (e.g., *IDI1* and *SC4MOL*).

25-HC has recently been shown to mediate anti-inflammatory activities via SREBP2 [30]. Here, we find miR-342-5p functions in a SREBP-dependent and -independent manner to regulate sterol biosynthesis and viral infection. Whether miR-342-5p has SREBP-related inflammatory functions remains to be fully investigated; however, the current study strongly supports the possibility that miR-342-5p might contribute to the regulation of inflammatory responses by targeting SREBP2 or the prenylation arm of the sterol pathway. In this context, a proinflammatory role for miR-342-5p in the enhancement of miR-155 expression has been described and attributed to the former miRNA targeting *BMPRI* (Entrez Gene: 12166) and *AKT1* (Entrez Gene: 11651) transcripts in macrophages [42]. In this study, we did not observe any notable alterations in *BMPRI* transcript synthesis or abundance during 8 h of IFN- γ BMDM treatment. We did, however, observe a small but significant reduction in *AKT1* abundance over time in IFN-stimulated macrophages. A knockdown of this transcript with a siRNA, however, had no effect on MCMV replication (S9E Fig).

Intriguingly, miR-342-5p targets multiple members of the sterol biosynthesis pathway including its master regulator SREBP2. The suppression of SREBP2 function results in a generalised decrease in sterol biosynthesis as well as a reduction in the abundance of key contributors to cholesterol homeostasis such as *INSIG1* (Entrez Gene: 231070; Fig 7C, S2 and S3 Figs) and *LDLR* (Entrez Gene: 16835; S8D Fig). Both DNA and RNA viruses require the sterol biosynthesis pathway for optimal growth capacity, and we have recently shown that an IFN-mediated suppression of sterol metabolic network activity is an integral part of the antiviral response [24]. Importantly, however, whilst the IFN regulation of SREBP2 is undoubtedly important, alternative mechanisms play a dominant role in the sterol-related antiviral responses we have observed to-date.

Notably, in this study we found miR-342-5p directly targets enzymes of both the proximal and distal arms of the sterol biosynthesis pathway—in particular *IDI1*. *IDI1* is an enzyme critical to protein prenylation and catalyses the isomerisation of the inactive carbon-carbon double bond of isopentenyl diphosphate (IPP) to generate an isomer dimethylallyl diphosphate (DMAPP). Prenylation is a post-translational modification enabling the membrane association of modified proteins and involves the covalent addition of prenyl lipids (e.g., farnesyl or geranylgeranyl) derived from mevalonic acid to conserved residues at the C-terminus of proteins. This process is integral to host intracellular protein trafficking, leukocyte chemotaxis, and phagocytosis and has also been implicated in inflammatory cytokine production [53–56]. In this connection, we have recently highlighted the importance of the mevalonate-isoprenoid branch point to the antiviral effects of 25-HC [24].

The replication of several viruses requires prenylation of host and/or virus proteins. For example, Hepatitis D virus requires prenylation of its large delta antigen for optimal virion morphogenesis, and prenylation inhibitors have shown promise in the treatment of this pathogen [48]. Further, Hepatitis C virus requires the geranylgeranylated host protein FBL2 for

replication and respiratory syncytial virus (RSV) F glycoprotein binds to the prenylated host protein RHOA enabling membrane fusion [57,58]. Here, we found that siRNA knockdown of IDI1 inhibits MCMV replication in pMEF [18] and, most importantly, the inhibitory effects of miR-342-5p on MCMV could be partially rescued via exogenous administration of GGOH and, to a lesser degree, MVA to cells. This points to an involvement of the sterol pathway prenylation branch point in the anti-MCMV functionality of miR-342-5p. However, the mode of action of miR-342-5p on the prenylation branch point remains to be determined. For Influenza virus, the inhibitory effects of miR-342-5p were partially rescued via the administration of exogenous MEV, FOH, SQL and, to a lesser extent, GGOH to cells. Thus, different targeting mechanisms of miR-342-5p facilitate broad antiviral effector functions.

The metabolic rescue data described above is crucial to our findings. It confirms sterol pathway targeting is essential to the antiviral effects of miR-342-5p we observed and highlights that whilst many viruses depend on the sterol metabolic network for their replication, unrelated virus types may exploit subtly different aspects of this pathway for their replication cycle. As a result, “multihit” targeting of a single pathway by miR-342-5p has beneficial outcomes for its broad antiviral effects.

Studies supporting a role for antiviral RNA-mediated interference in mammalian systems are beginning to emerge; however, the IFN response is still considered the pre-eminent mechanism by which mammalian cells resist viral infection [59–61]. Specific targeting of viral transcripts by host miRNAs has been reported, e.g., for hepatitis C virus (HCV) [6]. However, this is unlikely to represent a major mechanism for restricting viruses, as they are predisposed to the emergence of escape mutants [62]. In contrast, IFN-elicited miRNAs targeting cellular pathways required for virus replication arguably provide more robust and durable effects against a broad spectrum of viruses. Here, to our knowledge, we identify for the first time an IFN-induced host-targeting miRNA that elicits broad antiviral effects. Whilst we cannot preclude the possibility that miR-342-5p has a direct effect on viral RNA expression, the breadth of viruses repressed by this miRNA (both DNA and RNA) argues against a direct targeting of viral transcripts. Although miR-342-5p has been shown to specifically inhibit coxsackie B virus [51], our work emphasises the multifunctional, dual potential of this IFN-regulated antiviral miRNA.

In conclusion, the multihit targeting of sterol synthesis and antiviral effects mediated by miR-342-5p represent a new arm of the IFN-induced cell-autonomous immune response to viral infection and a new mechanistic link between lipid metabolism and the very early innate immune response. In this regard, miR-342-5p and its sterol pathway targets provide the foundation for future therapeutic exploitation, and this study highlights a general principle for blockade of infection.

Materials and Methods

Mice

C57BL/6 mice were housed in the specific pathogen-free animal facility at the University of Edinburgh. BALB/c mice were housed in the specific pathogen-free animal facility at the Institut d'Investigacions Biomèdiques August Pi i Sunyer, Barcelona, Spain. CH25H^{-/-} (B6.129S6-Ch25h^{tm1Rus}/J) mice were purchased from Charles River (Margate, United Kingdom) and housed in the specific pathogen-free animal facility at the University of Edinburgh. All procedures were carried out under project and personal licences approved by the Secretary of State for the Home Office, under the United Kingdom's 1986 Animals (Scientific Procedures) Act and the Local Ethical Review Committee at Edinburgh University. All procedures involving animals and their care in Spain were approved by the Ethics Committee (protocol

number CEEA 308/12) of the University of Barcelona and were conducted in compliance with institutional guidelines as well as with national (Generalitat de Catalunya decree 214/1997, DOGC 2450) and international (Guide for the Care and Use of Laboratory Animals, National Institutes of Health, 85–23, 1985) laws and policies.

Cell Propagation and Culture

BMDMs were isolated and differentiated with CSF-1 derived from L929 cells. Details of all cell culture conditions are provided in [S1 Methods](#).

Treatment of Cells with IFN or LPS

Unless otherwise stated, murine recombinant IFN gamma (IFN- γ) (Perbio Science) and IFN- β (Stratech, UK) were added to cells at a final concentration of 10 U/ml or 25 U/ml, respectively. For experiments investigating the effect of the miR-342-5p inhibitor on SREBF2 transcript abundance in BMDM, subconfluent macrophages were transfected for 24 h and then treated with 2.5 ng/ml murine recombinant IFNG (Life Technologies) for 24 h. Lipopolysaccharides from *Escherichia coli* 026:B6 (Sigma-Aldrich, UK) were reconstituted in SPBS (1 mg/ml) and added to BMDM at a final concentration of 100 ng/ml.

Reporter Viruses and Viral Plaque Assay

The GFP-encoding MCMV (MCMV-GFP) has been previously described [63]. HCMV-GFP (AD169-GFP) has been previously described [64]. HSV-1-eGFP (C12) was propagated and titred by plaque assay in Vero cells. A/WSN/33 (H1N1) influenza virus was propagated and titred in MDCK cells. Growth and titration conditions are provided in [S1 Methods](#).

GFP-Virus growth

MiRNA mimics or siRNA were transfected into cells using DharmaFECT 1 (Thermo Fisher Scientific). After 48 h or 72 h, cells were infected with virus and growth was measured using a POLARstar OPTIMA plate reader (BMG Labtech) according to manufacturers' recommendations. See [S1 Methods](#).

BMDM IFN- γ Treatment, RNA Labelling and Isolation

Incorporation of 4-thiouridine (Sigma) into newly-transcribed RNA was undertaken as described by Dölken et al. [65]. See [S1 Methods](#) for further information.

Newly Transcribed RNA Labelling for Microarray Analysis

Processing of ntRNA samples (100 ng) for hybridisation to Affymetrix Mouse Gene 1.0 ST arrays was undertaken according to manufacturer's instructions (Affymetrix). Hybridisation, washing, staining, and scanning of the arrays were also undertaken following standard Affymetrix protocols. After scanning and data capture, open-source R-based software "Bioconductor" was used to implement all quality control and statistical analyses. See [S1 Methods](#) for further information.

Q-RT-PCR Analysis of Cholesterol Pathway Genes

Total RNA was extracted from cells with RNeasy Mini kit (QIAGEN). Quantitative gene-expression analyses were then performed using Roche UPL reagents, IDT PrimeTime (IDT, United States) assays or Taqman primer probe sets (Applied Biosystems). Expression of target

genes was normalized to Actb unless otherwise stated. See [S1 Methods](#) for further detailed information.

PCR Analysis of Mature miRNA

Total RNA from in vitro tissue culture experiments was isolated using a Qiagen miRNeasy kit according to manufacturer's recommendations (Qiagen, US). MiRNA expression analyses were then performed using reagents and assays from Quanta Biosciences, US as per manufacturers recommendations. See [S1 Methods](#) for further detailed information.

PCR Data Analysis

Stratagene MXPro software was used to analyse the data. Threshold determinations and differences in transcript abundance relative to controls were automatically performed by software for each reaction.

Transfection of Mimics, Inhibitors, and siRNA

MicroRNA mimics, control miRNAs and siRNA were purchased from Dharmacon RNAi Technologies, Thermo Fisher Scientific (Lafayette, USA). Mimics and siRNA were transfected into cells using DharmaFECT 1 (Thermo Fisher Scientific). ZEN-AMO and 2'OMe/LNA-PS miRNA inhibitors were obtained from IDT and for NIH/3T3 cells were transfected using DharmaFECT 1 [66]. For BMDM or RAW cells, miRNA mimics or inhibitors were transfected (final concentration 25 nM unless otherwise stated) into cells using Viromer Blue (Lipocalyx, Germany) as per manufacturers recommendations. See [S1 Methods](#) for further information.

Administration of Small RNAs to Mice

For in vivo experiments, miRNA were administered by an intraperitoneal injection route as previously described [67]. For infection, mice were injected with 1×10^6 PFU MCMV in SPBS on day 3 of the experiment. Mock-infected animals were injected in an identical manner with SPBS only. Tissues were collected and snap frozen for subsequent analysis of MCMV titre. See [S1 Methods](#) for further information.

Transfection of 3'UTR Luciferase Reporter Plasmids and miRNA

Regions from the 3'UTR of the gene were synthesized and subcloned into the 3'UTR MCS of the psiCheck2 renilla luciferase by Eurofins MWG Operon (Ebersberg, Germany). MiRNA mimics or controls were reverse transfected into cells with either wild-type or mutant luciferase reporter in DharmaFECT DUO. After 24 h, luciferase expression in the transfected cells was measured using a dual-luciferase reporter assay kit (Promega, UK). See [S1 Methods](#) for further information.

Treatment of Cells with 25-HC or (2-Hydroxypropyl)- β -Cyclodextrin

25-HC (Sigma, H1015) was dissolved in 100% Ethanol (1000x stock, 20 mM) and stored at -20°C under argon in 2 ml opaque tubes with gasket screw-top lids. The powder (2-Hydroxypropyl)- β -cyclodextrin (H β CD) (Sigma, H107) was dissolved in medium at 37°C just before use.

Measurement of Total Cholesterol Concentration by Enzymatic Assay

Cholesterol was extracted from cells using Chloroform/Methanol protocol detailed in [S1 Methods](#). Total Cholesterol quantitation was carried out using an Amplex Red Cholesterol Assay Kit as per manufacturers recommendations (Invitrogen, UK).

Analysis of Sterols by Mass Spectrometry

Sterols and oxysterols were analysed using liquid chromatography—mass spectrometry (LC-MS) on an Orbitrap Elite (ThermoFisher) operated as described previously [68]. Sterols and oxysterols were identified by comparison of m/z , retention time and MSⁿ fragmentation with reference standards. Quantification was by stable isotope-dilution. See [S1 Methods](#) for further information.

Computational Prediction of miRNA Target Binding Sites in Sterol-Related Genes

For the prediction of potential microRNA targets in genomic 3' UTR regions the database TargetScan was used [69]. For the prediction of microRNA targets in 5'UTR, coding regions and 3'UTRs the database miRWalk was used [70].

Computational Prediction of miRNA Target Binding Sites in Virus Genes

MiRanda version 3.0 was used to scan viral coding sequences for predicted miRNA binding sites. The viral coding sequences were obtained from GenBank via NCBI for the following accessions: MCMV (NC_004065.1), HCMV (FJ527563.1), HSV1 (X14112.1), and influenza A (A/WSN/33 (H1N1): X14112.1, CY010795.1, CY010794.1, CY010793.1, CY010788.1, CY010791.1, CY010790.1, CY010789.1 and CY010792.1). MiRanda results were extracted as key-value pairs and sorted according to total score and free energy. See [S1 Methods](#) for further information.

Computational Prediction of Transcription Factor Binding Sites in Evl Promoter from Human and Mouse

To analyse and predict potential Stat1, Irf1 and Irf9 binding sites in the promoter of Human and Mouse EVL/Evl, the open source software Toucan was used [71]. See [S1 Methods](#) for further information.

Chromatin Immunoprecipitation and PCR Analysis of Promoter Sequence Enrichment

BMDM were treated with Ifn- γ (10 U/ml) for 2, 6, and 24 h, fixed and then Chromatin immunoprecipitation (ChIP) was performed as described previously [72]. Primers for the amplification of promoter regions from EVL and the positive control gene CXCL10 were designed using PrimerBLAST and are provided in [S5 Fig](#). Quantitative-PCR using SYBR-green incorporation (Quanta PerfeCTa SYBR Green FastMix, Low ROX) was used to analyse enrichment of sequences relative to input DNA. See [S1 Methods](#) for detailed information.

Analysis of EVL Promoter Activation

A 421-bp region from the Human EVL promoter containing three predicted IFN-activated transcription factor binding sites was synthesized and subcloned into the MCS of the pGL4.1 luciferase plasmid by Eurofins MWG Operon (Ebersberg, Germany). In parallel, corresponding mutants of each individual site (designated: ISRE, proximal Irf7 and distal Irf7) and a mutant in which all predicted sites were mutated (designated: All) were produced. Promoter activation by type 1 IFN was then tested as described in [S1 Methods](#).

Metabolite and LXR Agonist Treatment of Cells

After washing, normal medium containing vehicle (Ethanol) or GGOH (Sigma G3278), Mevalonolactone (MEV) (Sigma M4667), FOH (Sigma F203), or SQL (Sigma S3626) was added to the infected wells. T0901317 (Tocris Bioscience, Bristol, UK) was resuspended in 100% ethanol (5 mM stock), diluted in normal medium (50 nM final concentration) and cells were treated for 18 h.

Statistical Analyses

Unless otherwise stated, a two-sample Welch *t* test was used to test statistical significance of results in Microsoft Excel. Prior to parametric testing, a Shapiro-Wilk test was undertaken in R to confirm normal distribution of data. Statistical testing of Q-RT-PCR data from independent experiments (normalised to housekeeping gene ACTB) was undertaken using a one-sample *t* test in Microsoft Excel. Experimental group sizes (where *n* = number of independent biological samples per group) are stated in figure legends. Statistical analyses of growth curves was undertaken using a permutation based approach using a browser-based implementation of the “compareGrowthCurves” function originally developed for the statmod software package for R (<http://bioinf.wehi.edu.au/software/compareCurves>).

Cholesterol Biosynthesis Model

The cholesterol biosynthesis model used in this study was derived from a comprehensive consensus Systems Biology Graphical Notation diagram of the regulation and feedback of cholesterol metabolism [28].

Supporting Information

S1 Data. Numerical values underlying summary data displayed in the figures.
(XLSX)

S1 Fig. Labelling and isolation of newly transcribed RNA.
(EPS)

S2 Fig. Ifng down-regulates synthesis of sterol pathway transcripts. Upper Graphs: Sequential analysis of sterol pathway-related transcript synthesis (every 30 min) in IFN- γ -treated BMDM (relative to mock). Log₂ fold change values were calculated by subtracting the Mock from the IFN- γ -treated Log₂ scale signal value. Lower graphs: Sequential analysis of *SREBF1*, *SREBF2*, *INSIG1*, *INSIG2*, and *SCAP* transcript synthesis (every 30 min) in IFN- γ -treated BMDM (relative to mock). Log₂ fold change values were calculated as described for (B).
(EPS)

S3 Fig. Ifng down-regulates abundance of sterol pathway transcripts. Upper graphs: Abundance of sterol biosynthesis pathway-related transcripts in IFN- γ -treated BMDM (relative to mock). Log₂ fold change values were calculated by subtracting the Mock from the IFN- γ treated Log₂ scale signal value. Lower graphs: Abundance of *SREBF1*, *SREBF2*, *INSIG1*, *INSIG2*, and *SCAP* transcripts in IFN- γ -treated BMDM (relative to mock). Log₂ fold change values were calculated as described for (A).
(EPS)

S4 Fig. IFN up-regulates the expression of EVL/ miR-342. (A) IFN- γ or - β effects on mature miR-342-5p abundance in BMDM 7 h or 24 h after treatment. Values are normalised to mock treated cells. Data are mean \pm SEM (*n* = 3), * *p* \leq 0.05. (B) IFN- γ or - β effects on mature

miR-155 abundance in BMDM 7 h or 24 h after treatment. Values are normalised to mock treated cells. Data are mean \pm SEM ($n = 3$), * $p \leq 0.05$, ** $p \leq 0.01$. (C) *EVL* and pri/pre-miR-342 RNA transcript synthesis and abundance in BMDM following IFN- γ stimulation (10 U/ml). (D) *EVL* transcript synthesis (relative to Mock infected cells) in wild-type or TYK2^{-/-} BMDM 60 to 90 and 360 to 390 min after MCMV infection. (E) Pre-miR-342 synthesis (relative to Mock infected cells) in wild-type or TYK2^{-/-} BMDM 60 to 90 and 360 to 390 min after MCMV infection. (EPS)

S5 Fig. IFN directly regulates *EVL*/ miR-342 transcription. (A) Predicted ISRE and IRF7 binding sites in *EVL* promoter derived from the software Toucan. (B) Chromatin immunoprecipitation analysis of STAT1 and IRF1 transcription factor binding to *ACTB* (negative control), *CXCL10* (positive control) or the *EVL* promoter in BMDM cultured with IFN- γ (10 U/ml) for 2 h (top) or 24 h (bottom). Data are representative of multiple independent experiments. (C) Primer sequences used to amplify murine *ACTB*, *CXCL10* or *EVL* promoter DNA isolated by Chromatin Immunoprecipitation. (EPS)

S6 Fig. *SREBF2* siRNA and miR-342-5p effects on cholesterol pathway transcript abundance. *SREBF2* siRNA (upper panel) and miR-342-5p (lower panel) effects on cholesterol pathway transcript abundance in pMEF. Data are mean \pm SEM ($n = 6$), ** $p \leq 0.01$. (EPS)

S7 Fig. MiR-342-5p targets *Srebf2* and miR-33 and regulates intracellular cholesterol by multiple mechanisms. (A and B) Effects of miR-342-5p on relative *SREBF1* or *SREBF2* RNA abundance in mouse and human cells. * $p \leq 0.05$, ** $p \leq 0.01$ ($n > 2$). (C) IFN- γ or - β effects on mature miR-33 abundance in BMDM 7 h or 24 h after treatment. Values are normalised to mock treated cells. Data are mean \pm SEM ($n = 3$), * $p \leq 0.05$, ** $p \leq 0.01$. (D) Schematic showing relationship between LXR regulated transcripts *ABCA1* and *ABCG1*, miR-33 and miR-342-5p. (E) MiR-342-5p effect on miR-33-5p and miR-33-3p abundance in resting or T0901317 treated BMDM. (EPS)

S8 Fig. MiR-342-5p regulates intracellular cholesterol by multiple mechanisms. (A) Quantitation of miR-342-5p effects on total intracellular cholesterol in NIH/3T3 fibroblasts. Data are mean \pm SEM ($n = 3$) * $p \leq 0.05$. (B) Quantitation of miR-342-5p inhibitor effects on total intracellular cholesterol in NIH/3T3 fibroblasts. Data are mean \pm SEM ($n = 2$). (C) Reduction of Mouse *DHCR7* 3'UTR reporter expression by miR-342-5p. Data are mean \pm SEM ($n = 3$). (D) MiR-342-5p reduces LDLR mRNA abundance in NIH/3T3 cells. Data are mean \pm SEM ($n = 3$), ** $p < 0.01$. (EPS)

S9 Fig. MiR-342-5p regulation of MCMV in vitro and in vivo. (A) Effects of IFN-regulated miRNA on MCMV-GFP replication. Data are normalised to values obtained with non-targeting siRNA and are mean \pm SEM ($n = 2$). (B) Effects of IFN- γ treatment (100 pg/ml for 6 h) on MCMV-GFP replication in negative control transfected NIH/3T3 cells. $N = 3$, * $p \leq 0.05$. (C and D) MCMV titre (4 dpi) in Spleen (C) and Liver (D) of BALB/c mice treated with 10 μ g or 40 μ g miR-342-5p or control miRNA. Data are titre in organ of individual mouse ($n = 6$). * $p \leq 0.05$, ** $p \leq 0.01$. (E) Effects of *AKT1* siRNA miRNA on MCMV-GFP replication. Data are normalised to values obtained with nontargeting siRNA and are mean \pm SEM ($n = 3$). ** $p \leq 0.01$. (F) Effects of *HMGCR*, *IDI1*, *SC4MOL*, and *DHCR7* siRNA on the abundance of

their respective target RNAs in MEF transfected for 48 h. Data are mean \pm SEM ($n = 3$).

$**p \leq 0.01$.

(EPS)

S10 Fig. MiR-342-5p does not affect cell viability. (A and B) Analysis of miR-342-5p effects on cell viability. Primary MEFs (A), 3T3, MRC-5 and HeLa (B) cells were transfected with the indicated doses of control siRNA/ miRNA or miR-342-5p mimics 48 h before analysis using the CellTiter-Blue viability assay. (C) Schematic showing sterol biosynthesis pathway entry points for metabolites used in metabolic rescue experiments.

(EPS)

S11 Fig. Schematic showing IFN-activated 25-HC and miR-342-5p pathways regulating sterol metabolism.

(EPS)

S1 Methods. Extended descriptions of materials and methods.

(DOCX)

S1 Table. MiR-342-5p is predicted to target the 3'UTR of SREBF2 and multiple cholesterol pathway transcripts. The database TargetScan was queried using mouse Entrez gene IDs for cholesterol biosynthesis pathway members. Target predictions for mouse miRNAs were then tabulated.

(DOCX)

S2 Table. MiR-342- 5p is predicted to target Herpesvirus but not Influenza virus transcripts. MiRanda version 3.0 was used to scan viral coding sequences for predicted miRNA binding sites. Results were extracted as key-value pairs and sorted according to total score and free energy.

(DOCX)

Acknowledgments

We wish to thank Caroline Friedel, Diwakar Santhakumar, Amy Buck, Birgit Ströbl, Finn Grey, Peter Phelan, Wendy Bickmore, and Tim Osborne for their help, comments, and sharing of reagents. We are grateful to David Russell for the CH25H^{-/-} mice. We also wish to thank Alan Ross and Marie Craigon for technical assistance.

Author Contributions

Conceived and designed the experiments: KAR PG MB. Performed the experiments: KAR WYH PJC EY YW WJG KM MY MMP SJG AA HL ST. Analyzed the data: KAR TF SW WJG AJE PG. Contributed reagents/materials/analysis tools: LD JH KAL MAB AJE MMP. Wrote the paper: KAR PG.

References

1. Karlikow M, Goic B, Saleh M. RNAi and antiviral defense in Drosophila: Setting up a systemic immune response. *Developmental and Comparative Immunology*. 2014; 42(1):85–92. doi: [10.1016/j.dci.2013.05.004](https://doi.org/10.1016/j.dci.2013.05.004) WOS:000328174200010. PMID: [23684730](https://pubmed.ncbi.nlm.nih.gov/23684730/)
2. Szittya G, Burgyan J, Cullen B. RNA Interference-Mediated Intrinsic Antiviral Immunity in Plants. *Intrinsic Immunity*. 2013; 371:153–81. doi: [10.1007/978-3-642-37765-5_6](https://doi.org/10.1007/978-3-642-37765-5_6) WOS:000330591400007.
3. Schoggins JW. Interferon-stimulated genes: roles in viral pathogenesis. *Curr Opin Virol*. 2014; 6:40–6. doi: [10.1016/j.coviro.2014.03.006](https://doi.org/10.1016/j.coviro.2014.03.006) PMID: [24713352](https://pubmed.ncbi.nlm.nih.gov/24713352/); PubMed Central PMCID: PMCPMC4077717.

4. Sadler A, Williams B. Interferon-inducible antiviral effectors. *Nature Reviews Immunology*. 2008; 8(7):559–68. doi: [10.1038/nri2314](https://doi.org/10.1038/nri2314) WOS:000257089200016. PMID: [18575461](https://pubmed.ncbi.nlm.nih.gov/18575461/)
5. Santhakumar D, Forster T, Laqtom NN, Fragkoudis R, Dickinson P, Abreu-Goodger C, et al. Combined agonist-antagonist genome-wide functional screening identifies broadly active antiviral microRNAs. *Proceedings of the National Academy of Sciences of the United States of America*. 2010; 107(31):13830–5. Epub 2010/07/21. doi: [10.1073/pnas.1008861107](https://doi.org/10.1073/pnas.1008861107) PMID: [20643939](https://pubmed.ncbi.nlm.nih.gov/20643939/); PubMed Central PMCID: PMC2922253.
6. Pedersen IM, Cheng G, Wieland S, Volinia S, Croce CM, Chisari FV, et al. Interferon modulation of cellular microRNAs as an antiviral mechanism. *Nature*. 2007; 449(7164):919–22. Epub 2007/10/19. doi: [10.1038/nature06205](https://doi.org/10.1038/nature06205) PMID: [17943132](https://pubmed.ncbi.nlm.nih.gov/17943132/); PubMed Central PMCID: PMC2748825.
7. Cazalla D, Yario T, Steitz J. Down-Regulation of a Host MicroRNA by a Herpesvirus saimiri Noncoding RNA. *Science*. 2010; 328(5985):1563–6. doi: [10.1126/science.1187197](https://doi.org/10.1126/science.1187197) WOS:000278859200048. PMID: [20558719](https://pubmed.ncbi.nlm.nih.gov/20558719/)
8. Fu Y, Liu X, Li X, Shen Z, Yang B, Wu C, et al. MicroRNA miR-21 Attenuates Human Cytomegalovirus Replication in Neural Cells by Targeting Cdc25a. *Journal of Virology*. 2015; 89(2):1070–82. doi: [10.1128/JVI10740-14](https://doi.org/10.1128/JVI10740-14) WOS:000347178900016. PMID: [25378484](https://pubmed.ncbi.nlm.nih.gov/25378484/)
9. Reinsbach S, Nazarov P, Philippidou D, Schmitt M, Wienecke-Baldacchino A, Muller A, et al. Dynamic regulation of microRNA expression following interferon gamma-induced gene transcription. *Rna Biology*. 2012; 9(7):978–89. doi: [10.4161/ma.20494](https://doi.org/10.4161/ma.20494) WOS:000307078700009. PMID: [22767256](https://pubmed.ncbi.nlm.nih.gov/22767256/)
10. van der Ree M, van der Meer A, de Bruijne J, Maan R, van Vliet A, Welzel T, et al. Long-term safety and efficacy of microRNA-targeted therapy in chronic hepatitis C patients. *Antiviral Research*. 2014; 111:53–9. doi: [10.1016/j.antiviral.2014.08.015](https://doi.org/10.1016/j.antiviral.2014.08.015) WOS:000345185100009. PMID: [25218783](https://pubmed.ncbi.nlm.nih.gov/25218783/)
11. Wang P, Hou J, Lin L, Wang C, Liu X, Li D, et al. Inducible microRNA-155 Feedback Promotes Type I IFN Signaling in Antiviral Innate Immunity by Targeting Suppressor of Cytokine Signaling 1. *Journal of Immunology*. 2010; 185(10):6226–33. doi: [10.4049/jimmunol.1000491](https://doi.org/10.4049/jimmunol.1000491) WOS:000283848000061.
12. Ghazal P, Armas J, Garcia-Ramirez J, Kurz S, Angulo A. Viruses: Hostages to the cell. *Virology*. 2000; 275(2):233–7. doi: [10.1006/viro.2000.0553](https://doi.org/10.1006/viro.2000.0553) WOS:000089697900002. PMID: [10998321](https://pubmed.ncbi.nlm.nih.gov/10998321/)
13. Schoggins JW, Randall G. Lipids in innate antiviral defense. *Cell Host Microbe*. 2013; 14(4):379–85. Epub 2013/10/22. doi: [10.1016/j.chom.2013.09.010](https://doi.org/10.1016/j.chom.2013.09.010) PMID: [24139397](https://pubmed.ncbi.nlm.nih.gov/24139397/)
14. del Real G, Jimenez-Baranda S, Mira E, Lacalle RA, Lucas P, Gomez-Mouton C, et al. Statins inhibit HIV-1 infection by down-regulating Rho activity. *J Exp Med*. 2004; 200(4):541–7. Epub 2004/08/18. doi: [10.1084/jem.20040061](https://doi.org/10.1084/jem.20040061) PMID: [15314078](https://pubmed.ncbi.nlm.nih.gov/15314078/); PubMed Central PMCID: PMC2211926.
15. Potenza L, Frascaroli G, Grigioni F, Lazzarotto T, Magnani G, Tomasi L, et al. Hydroxymethyl-glutaryl coenzyme a reductase inhibition limits cytomegalovirus infection in human endothelial cells. *Circulation*. 2004; 109(4):532–6. Epub 2004/01/28. doi: [10.1161/01.CIR.0000109485.79183.81](https://doi.org/10.1161/01.CIR.0000109485.79183.81) PMID: [14744969](https://pubmed.ncbi.nlm.nih.gov/14744969/).
16. Ye J, Wang C, Sumpter R Jr., Brown MS, Goldstein JL, Gale M Jr. Disruption of hepatitis C virus RNA replication through inhibition of host protein geranylgeranylation. *Proceedings of the National Academy of Sciences of the United States of America*. 2003; 100(26):15865–70. Epub 2003/12/12. doi: [10.1073/pnas.2237238100](https://doi.org/10.1073/pnas.2237238100) PMID: [14668447](https://pubmed.ncbi.nlm.nih.gov/14668447/); PubMed Central PMCID: PMC307659.
17. Yu Y, Maguire TG, Alwine JC. Human cytomegalovirus infection induces adipocyte-like lipogenesis through activation of sterol regulatory element binding protein 1. *Journal of virology*. 2012; 86(6):2942–9. Epub 2012/01/20. doi: [10.1128/JVI.06467-11](https://doi.org/10.1128/JVI.06467-11) PMID: [22258239](https://pubmed.ncbi.nlm.nih.gov/22258239/); PubMed Central PMCID: PMC3302344.
18. Blanc M, Hsieh WY, Robertson KA, Kropp KA, Forster T, Shui G, et al. The transcription factor STAT-1 couples macrophage synthesis of 25-hydroxycholesterol to the interferon antiviral response. *Immunity*. 2013; 38(1):106–18. doi: [10.1016/j.immuni.2012.11.004](https://doi.org/10.1016/j.immuni.2012.11.004) PMID: [23273843](https://pubmed.ncbi.nlm.nih.gov/23273843/); PubMed Central PMCID: PMC3556782.
19. Rodgers M, Villareal V, Schaefer E, Peng L, Corey K, Chung R, et al. Lipid Metabolite Profiling Identifies Desmosterol Metabolism as a New Antiviral Target for Hepatitis C Virus. *Journal of the American Chemical Society*. 2012; 134(16):6896–9. doi: [10.1021/ja207391q](https://doi.org/10.1021/ja207391q) WOS:000303139800003. PMID: [22480142](https://pubmed.ncbi.nlm.nih.gov/22480142/)
20. Im SS, Yousef L, Blaschitz C, Liu JZ, Edwards RA, Young SG, et al. Linking lipid metabolism to the innate immune response in macrophages through sterol regulatory element binding protein-1a. *Cell metabolism*. 2011; 13(5):540–9. Epub 2011/05/03. doi: [10.1016/j.cmet.2011.04.001](https://doi.org/10.1016/j.cmet.2011.04.001) PMID: [21531336](https://pubmed.ncbi.nlm.nih.gov/21531336/); PubMed Central PMCID: PMC3090630.
21. Kidani Y, Elsaesser H, Hock MB, Vergnes L, Williams KJ, Argus JP, et al. Sterol regulatory element-binding proteins are essential for the metabolic programming of effector T cells and adaptive immunity. *Nature immunology*. 2013; 14(5):489–99. Epub 2013/04/09. doi: [10.1038/ni.2570](https://doi.org/10.1038/ni.2570) PMID: [23563690](https://pubmed.ncbi.nlm.nih.gov/23563690/); PubMed Central PMCID: PMC3652626.

22. Seo YK, Jeon TI, Chong HK, Biesinger J, Xie X, Osborne TF. Genome-wide localization of SREBP-2 in hepatic chromatin predicts a role in autophagy. *Cell metabolism*. 2011; 13(4):367–75. Epub 2011/04/05. doi: [10.1016/j.cmet.2011.03.005](https://doi.org/10.1016/j.cmet.2011.03.005) PMID: [21459322](https://pubmed.ncbi.nlm.nih.gov/21459322/); PubMed Central PMCID: PMC3086264.
23. Ogawa S, Lozach J, Benner C, Pascual G, Tangirala RK, Westin S, et al. Molecular determinants of crosstalk between nuclear receptors and toll-like receptors. *Cell*. 2005; 122(5):707–21. Epub 2005/09/07. doi: [10.1016/j.cell.2005.06.029](https://doi.org/10.1016/j.cell.2005.06.029) PMID: [16143103](https://pubmed.ncbi.nlm.nih.gov/16143103/); PubMed Central PMCID: PMC1430687.
24. Blanc M, Hsieh WY, Robertson KA, Watterson S, Shui G, Lacaze P, et al. Host Defense against Viral Infection Involves Interferon Mediated Down-Regulation of Sterol Biosynthesis. *PLoS Biol*. 2011; 9(3): e1000598. Epub 2011/03/17. doi: [10.1371/journal.pbio.1000598](https://doi.org/10.1371/journal.pbio.1000598) PMID: [21408089](https://pubmed.ncbi.nlm.nih.gov/21408089/); PubMed Central PMCID: PMC3050939.
25. Watterson S, Guerriero ML, Blanc M, Mazein A, Loewe L, Robertson KA, et al. A model of flux regulation in the cholesterol biosynthesis pathway: Immune mediated graduated flux reduction versus statin-like led stepped flux reduction. *Biochimie*. 2012. Epub 2012/06/06. doi: [10.1016/j.biochi.2012.05.024](https://doi.org/10.1016/j.biochi.2012.05.024) PMID: [22664637](https://pubmed.ncbi.nlm.nih.gov/22664637/).
26. Spann NJ, Garmire LX, McDonald JG, Myers DS, Milne SB, Shibata N, et al. Regulated accumulation of desmosterol integrates macrophage lipid metabolism and inflammatory responses. *Cell*. 2012; 151(1):138–52. Epub 2012/10/02. doi: [10.1016/j.cell.2012.06.054](https://doi.org/10.1016/j.cell.2012.06.054) PMID: [23021221](https://pubmed.ncbi.nlm.nih.gov/23021221/); PubMed Central PMCID: PMC3464914.
27. Liu SY, Aliyari R, Chikere K, Li G, Marsden MD, Smith JK, et al. Interferon-inducible cholesterol-25-hydroxylase broadly inhibits viral entry by production of 25-hydroxycholesterol. *Immunity*. 2013; 38(1):92–105. Epub 2013/01/01. doi: [10.1016/j.immuni.2012.11.005](https://doi.org/10.1016/j.immuni.2012.11.005) PMID: [23273844](https://pubmed.ncbi.nlm.nih.gov/23273844/); PubMed Central PMCID: PMC3698975.
28. Mazein A, Watterson S, Hsieh WY, Griffiths WJ, Ghazal P. A comprehensive machine-readable view of the mammalian cholesterol biosynthesis pathway. *Biochem Pharmacol*. 2013; 86(1):56–66. doi: [10.1016/j.bcp.2013.03.021](https://doi.org/10.1016/j.bcp.2013.03.021) PMID: [23583456](https://pubmed.ncbi.nlm.nih.gov/23583456/); PubMed Central PMCID: PMC3912678.
29. Sato R, Inoue J, Kawabe Y, Kodama T, Takano T, Maeda M. Sterol-dependent transcriptional regulation of sterol regulatory element-binding protein-2. *Journal of Biological Chemistry*. 1996; 271(43):26461–4. WOS:A1996VP23300005. PMID: [8900111](https://pubmed.ncbi.nlm.nih.gov/8900111/)
30. Reboldi A, Dang E, McDonald J, Liang G, Russell D, Cyster J. 25-Hydroxycholesterol suppresses interleukin-1-driven inflammation downstream of type I interferon. *Science*. 2014; 345(6197):679–84. doi: [10.1126/science.1254790](https://doi.org/10.1126/science.1254790) WOS:000339962800041. PMID: [25104388](https://pubmed.ncbi.nlm.nih.gov/25104388/)
31. Shibata N, Carlin A, Spann N, Saijo K, Morello C, McDonald J, et al. 25-Hydroxycholesterol Activates the Integrated Stress Response to Reprogram Transcription and Translation in Macrophages. *Journal of Biological Chemistry*. 2013; 288(50):35812–23. doi: [10.1074/jbc.M113.519637](https://doi.org/10.1074/jbc.M113.519637) WOS:000329814700019. PMID: [24189069](https://pubmed.ncbi.nlm.nih.gov/24189069/)
32. Rayner KJ, Suarez Y, Davalos A, Parathath S, Fitzgerald ML, Tamehiro N, et al. MiR-33 contributes to the regulation of cholesterol homeostasis. *Science*. 2010; 328(5985):1570–3. Epub 2010/05/15. sci-ence.1189862 [pii] doi: [10.1126/science.1189862](https://doi.org/10.1126/science.1189862) PMID: [20466885](https://pubmed.ncbi.nlm.nih.gov/20466885/).
33. Hu Z, Shen WJ, Kraemer FB, Azhar S. MicroRNAs 125a and 455 repress lipoprotein-supported steroidogenesis by targeting scavenger receptor class B type I in steroidogenic cells. *Molecular and cellular biology*. 2012; 32(24):5035–45. Epub 2012/10/10. doi: [10.1128/mcb.01002-12](https://doi.org/10.1128/mcb.01002-12) PMID: [23045399](https://pubmed.ncbi.nlm.nih.gov/23045399/); PubMed Central PMCID: PMC3510537.
34. Miller AM, Gilchrist DS, Nijjar J, Araldi E, Ramirez CM, Lavery CA, et al. MiR-155 has a protective role in the development of non-alcoholic hepatosteatosis in mice. *PLoS ONE*. 2013; 8(8):e72324. Epub 2013/08/31. doi: [10.1371/journal.pone.0072324](https://doi.org/10.1371/journal.pone.0072324) PMID: [23991091](https://pubmed.ncbi.nlm.nih.gov/23991091/); PubMed Central PMCID: PMC3749101.
35. Larsen L, Rosenstjerne MW, Gaarn LW, Bagge A, Pedersen L, Dahmcke CM, et al. Expression and localization of microRNAs in perinatal rat pancreas: role of miR-21 in regulation of cholesterol metabolism. *PLoS ONE*. 2011; 6(10):e25997. Epub 2011/10/25. doi: [10.1371/journal.pone.0025997](https://doi.org/10.1371/journal.pone.0025997) PMID: [22022489](https://pubmed.ncbi.nlm.nih.gov/22022489/); PubMed Central PMCID: PMC3191174.
36. Yang M, Liu W, Pellicane C, Sahyoun C, Joseph B, Gallo-Ebert C, et al. Identification of miR-185 as a regulator of de novo cholesterol biosynthesis and low density lipoprotein uptake. *Journal of Lipid Research*. 2014; 55(2):226–38. doi: [10.1194/jlr.M041335](https://doi.org/10.1194/jlr.M041335) WOS:000330535800007. PMID: [24296663](https://pubmed.ncbi.nlm.nih.gov/24296663/)
37. Ramirez CM, Rotllan N, Vlassov AV, Davalos A, Li M, Goedeke L, et al. Control of cholesterol metabolism and plasma high-density lipoprotein levels by microRNA-144. *Circulation research*. 2013; 112(12):1592–601. Epub 2013/03/23. doi: [10.1161/circresaha.112.300626](https://doi.org/10.1161/circresaha.112.300626) PMID: [23519695](https://pubmed.ncbi.nlm.nih.gov/23519695/).
38. O'Connell RM, Taganov KD, Boldin MP, Cheng G, Baltimore D. MicroRNA-155 is induced during the macrophage inflammatory response. *Proceedings of the National Academy of Sciences of the United States of America*. 2007; 104(5):1604–9. Epub 2007/01/24. doi: [10.1073/pnas.0610731104](https://doi.org/10.1073/pnas.0610731104) PMID: [17242365](https://pubmed.ncbi.nlm.nih.gov/17242365/); PubMed Central PMCID: PMC1780072.

39. Griffiths-Jones S, Saini HK, van Dongen S, Enright AJ. miRBase: tools for microRNA genomics. *Nucleic Acids Res.* 2008; 36:D154–8. Epub 2007/11/10. doi: [10.1093/nar/gkm952](https://doi.org/10.1093/nar/gkm952) PMID: [17991681](https://pubmed.ncbi.nlm.nih.gov/17991681/); PubMed Central PMCID: PMC2238936.
40. Wu C, Macleod I, Su AI. BioGPS and MyGene.info: organizing online, gene-centric information. *Nucleic Acids Res.* 2013; 41(Database issue):D561–5. Epub 2012/11/24. doi: [10.1093/nar/gks1114](https://doi.org/10.1093/nar/gks1114) PMID: [23175613](https://pubmed.ncbi.nlm.nih.gov/23175613/); PubMed Central PMCID: PMC3531157.
41. Ghani S, Riemke P, Schonheit J, Lenze D, Stumm J, Hoogenkamp M, et al. Macrophage development from hematopoietic stem cells requires PU.1 coordinated microRNA expression. *Blood.* 2011. Epub 2011/07/07. doi: [10.1182/blood-2011-02-335141](https://doi.org/10.1182/blood-2011-02-335141) PMID: [21730352](https://pubmed.ncbi.nlm.nih.gov/21730352/).
42. Wei Y, Nazari-Jahantigh M, Chan L, Zhu M, Heyll K, Corbalan-Campos J, et al. The microRNA-342-5p fosters inflammatory macrophage activation through an Akt1- and microRNA-155-dependent pathway during atherosclerosis. *Circulation.* 2013; 127(15):1609–19. Epub 2013/03/21. doi: [10.1161/CIRCULATIONAHA.112.000736](https://doi.org/10.1161/CIRCULATIONAHA.112.000736) PMID: [23513069](https://pubmed.ncbi.nlm.nih.gov/23513069/).
43. De Marchis ML, Ballarino M, Salvatori B, Puzzolo MC, Bozzoni I, Fatica A. A new molecular network comprising PU.1, interferon regulatory factor proteins and miR-342 stimulates ATRA-mediated granulocytic differentiation of acute promyelocytic leukemia cells. *Leukemia.* 2009; 23(5):856–62. Epub 2009/01/20. leu2008372 [pii] doi: [10.1038/leu.2008.372](https://doi.org/10.1038/leu.2008.372) PMID: [19151778](https://pubmed.ncbi.nlm.nih.gov/19151778/).
44. Li X, Chen YT, Jossion S, Mukhopadhyay NK, Kim J, Freeman MR, et al. MicroRNA-185 and 342 inhibit tumorigenicity and induce apoptosis through blockade of the SREBP metabolic pathway in prostate cancer cells. *PLoS ONE.* 2013; 8(8):e70987. Epub 2013/08/21. doi: [10.1371/journal.pone.0070987](https://doi.org/10.1371/journal.pone.0070987) PMID: [23951060](https://pubmed.ncbi.nlm.nih.gov/23951060/); PubMed Central PMCID: PMC3739799.
45. Kropp K, Robertson K, Sing G, Rodriguez-Martin S, Blanc M, Lacaze P, et al. Reversible Inhibition of Murine Cytomegalovirus Replication by Gamma Interferon (IFN-gamma) in Primary Macrophages Involves a Primed Type I IFN-Signaling Subnetwork for Full Establishment of an Immediate-Early Anti-viral State. *Journal of Virology.* 2011; 85(19):10286–99. doi: [10.1128/JVI.00373-11](https://doi.org/10.1128/JVI.00373-11) WOS:000296253900061. PMID: [21775459](https://pubmed.ncbi.nlm.nih.gov/21775459/)
46. Spann N, Glass C. Sterols and oxysterols in immune cell function. *Nature Immunology.* 2013; 14(9):893–900. doi: [10.1038/ni.2681](https://doi.org/10.1038/ni.2681) WOS:000323377700006. PMID: [23959186](https://pubmed.ncbi.nlm.nih.gov/23959186/)
47. Kapadia SB, Chisari FV. Hepatitis C virus RNA replication is regulated by host geranylgeranylation and fatty acids. *Proceedings of the National Academy of Sciences of the United States of America.* 2005; 102(7):2561–6. Epub 2005/02/09. doi: [10.1073/pnas.0409834102](https://doi.org/10.1073/pnas.0409834102) PMID: [15699349](https://pubmed.ncbi.nlm.nih.gov/15699349/); PubMed Central PMCID: PMC549027.
48. Bordier BB, Ohkanda J, Liu P, Lee SY, Salazar FH, Marion PL, et al. In vivo antiviral efficacy of prenylation inhibitors against hepatitis delta virus. *The Journal of clinical investigation.* 2003; 112(3):407–14. Epub 2003/08/05. doi: [10.1172/jci17704](https://doi.org/10.1172/jci17704) PMID: [12897208](https://pubmed.ncbi.nlm.nih.gov/12897208/); PubMed Central PMCID: PMC166292.
49. Liao Z, Cimaskasy LM, Hampton R, Nguyen DH, Hildreth JE. Lipid rafts and HIV pathogenesis: host membrane cholesterol is required for infection by HIV type 1. *AIDS research and human retroviruses.* 2001; 17(11):1009–19. Epub 2001/08/04. doi: [10.1089/088922201300343690](https://doi.org/10.1089/088922201300343690) PMID: [11485618](https://pubmed.ncbi.nlm.nih.gov/11485618/).
50. Bruce E, Digard P, Stuart A. The Rab11 Pathway Is Required for Influenza A Virus Budding and Filament Formation. *Journal of Virology.* 2010; 84(12):5848–59. doi: [10.1128/JVI.00307-10](https://doi.org/10.1128/JVI.00307-10) WOS:000277733900001. PMID: [20357086](https://pubmed.ncbi.nlm.nih.gov/20357086/)
51. Wang L, Qin Y, Tong L, Wu S, Wang Q, Jiao Q, et al. MiR-342-5p suppresses coxsackievirus B3 biosynthesis by targeting the 2C-coding region. *Antiviral research.* 2012; 93(2):270–9. Epub 2011/12/27. doi: [10.1016/j.antiviral.2011.12.004](https://doi.org/10.1016/j.antiviral.2011.12.004) PMID: [22197249](https://pubmed.ncbi.nlm.nih.gov/22197249/).
52. Lu H, Talbot S, Robertson K, Watterson S, Forster T, Roy D, et al. Rapid proteasomal elimination of 3-hydroxy-3-methylglutaryl-CoA reductase by interferon-gamma in primary macrophages requires endogenous 25-hydroxycholesterol synthesis. *Steroids.* 2015; 99:219–29. doi: [10.1016/j.steroids.2015.02.022](https://doi.org/10.1016/j.steroids.2015.02.022) WOS:000356985000016. PMID: [25759117](https://pubmed.ncbi.nlm.nih.gov/25759117/)
53. Thurnher M, Gruenbacher G, Nussbaumer O. Regulation of mevalonate metabolism in cancer and immune cells. *Biochimica Et Biophysica Acta-Molecular and Cell Biology of Lipids.* 2013; 1831(6):1009–15. doi: [10.1016/j.bbalip.2013.03.003](https://doi.org/10.1016/j.bbalip.2013.03.003) WOS:000319309800001.
54. Roberts A, Kim C, Zhen L, Lowe J, Kapur R, Petryniak B, et al. Deficiency of the hematopoietic cell-specific Rho family GTPase Rac2 is characterized by abnormalities in neutrophil function and host defense. *Immunity.* 1999; 10(2):183–96. doi: [10.1016/S1074-7613\(00\)80019-9](https://doi.org/10.1016/S1074-7613(00)80019-9) WOS:000078972200006. PMID: [10072071](https://pubmed.ncbi.nlm.nih.gov/10072071/)
55. Philips M. The perplexing case of the geranylgeranyl transferase-deficient mouse. *Journal of Clinical Investigation.* 2011; 121(2):510–3. doi: [10.1172/JCI45952](https://doi.org/10.1172/JCI45952) WOS:000286913800012. PMID: [21266773](https://pubmed.ncbi.nlm.nih.gov/21266773/)
56. Khan O, Ibrahim M, Jonsson I, Karlsson C, Liu M, Sjogren A, et al. Geranylgeranyltransferase type I (GGTase-I) deficiency hyperactivates macrophages and induces erosive arthritis in mice. *Journal of*

- Clinical Investigation. 2011; 121(2):628–39. doi: [10.1172/JCI43758](https://doi.org/10.1172/JCI43758) WOS:000286913800027. PMID: [21266780](https://pubmed.ncbi.nlm.nih.gov/21266780/)
57. Wang C, Gale M Jr., Keller BC, Huang H, Brown MS, Goldstein JL, et al. Identification of FBL2 as a geranylgeranylated cellular protein required for hepatitis C virus RNA replication. *Molecular cell*. 2005; 18(4):425–34. Epub 2005/05/17. doi: [10.1016/j.molcel.2005.04.004](https://doi.org/10.1016/j.molcel.2005.04.004) PMID: [15893726](https://pubmed.ncbi.nlm.nih.gov/15893726/).
58. Gower TL, Graham BS. Antiviral activity of lovastatin against respiratory syncytial virus in vivo and in vitro. *Antimicrobial agents and chemotherapy*. 2001; 45(4):1231–7. Epub 2001/03/21. doi: [10.1128/aac.45.4.1231-1237.2001](https://doi.org/10.1128/aac.45.4.1231-1237.2001) PMID: [11257039](https://pubmed.ncbi.nlm.nih.gov/11257039/); PubMed Central PMCID: PMC90448.
59. Turner M, Galloway A, Vigorito E. Noncoding RNA and its associated proteins as regulatory elements of the immune system. *Nature Immunology*. 2014; 15(6):484–91. doi: [10.1038/ni.2887](https://doi.org/10.1038/ni.2887) WOS:000336419300002. PMID: [24840979](https://pubmed.ncbi.nlm.nih.gov/24840979/)
60. Maillard P, Ciaudo C, Marchais A, Li Y, Jay F, Ding S, et al. Antiviral RNA Interference in Mammalian Cells. *Science*. 2013; 342(6155):235–8. doi: [10.1126/science.1241930](https://doi.org/10.1126/science.1241930) WOS:000325475200042. PMID: [24115438](https://pubmed.ncbi.nlm.nih.gov/24115438/)
61. Li Y, Lu J, Han Y, Fan X, Ding S. RNA Interference Functions as an Antiviral Immunity Mechanism in Mammals. *Science*. 2013; 342(6155):231–4. doi: [10.1126/science.1241911](https://doi.org/10.1126/science.1241911) WOS:000325475200041. PMID: [24115437](https://pubmed.ncbi.nlm.nih.gov/24115437/)
62. Heiss B, Maximova O, Thach D, Speicher J, Pletnev A. MicroRNA Targeting of Neurotropic Flavivirus: Effective Control of Virus Escape and Reversion to Neurovirulent Phenotype. *Journal of Virology*. 2012; 86(10):5647–59. doi: [10.1128/JVI.07125-11](https://doi.org/10.1128/JVI.07125-11) WOS:000303787100022. PMID: [22419812](https://pubmed.ncbi.nlm.nih.gov/22419812/)
63. Angulo A, Ghazal P, Messerle M. The major immediate-early gene ie3 of mouse cytomegalovirus is essential for viral growth. *Journal of virology*. 2000; 74(23):11129–36. Epub 2000/11/09. PMID: [11070009](https://pubmed.ncbi.nlm.nih.gov/11070009/); PubMed Central PMCID: PMC113196.
64. Borst E, Messerle M. Development of a cytomegalovirus vector for somatic gene therapy. *Bone Marrow Transplant*. 2000; 25 Suppl 2:S80–2. PMID: [10933196](https://pubmed.ncbi.nlm.nih.gov/10933196/).
65. Dolken L, Ruzsics Z, Radle B, Friedel CC, Zimmer R, Mages J, et al. High-resolution gene expression profiling for simultaneous kinetic parameter analysis of RNA synthesis and decay. *RNA*. 2008; 14(9):1959–72. Epub 2008/07/29. doi: [10.1261/rna.1136108](https://doi.org/10.1261/rna.1136108) [pii] doi: [10.1261/rna.1136108](https://doi.org/10.1261/rna.1136108) PMID: [18658122](https://pubmed.ncbi.nlm.nih.gov/18658122/); PubMed Central PMCID: PMC2525961.
66. Lennox K, Owczarzy R, Thomas D, Walder J, Behlke M. Improved Performance of Anti-miRNA Oligonucleotides Using a Novel Non-Nucleotide Modifier. *Molecular Therapy-Nucleic Acids*. 2013; 2. doi: [10.1038/mtna.2013.46](https://doi.org/10.1038/mtna.2013.46) WOS:000332467100005.
67. Lundberg P, Yang HJ, Jung SJ, Behlke MA, Rose SD, Cantin EM. Protection against TNFalpha-dependent liver toxicity by intraperitoneal liposome delivered DsiRNA targeting TNFalpha in vivo. *J Control Release*. 2012; 160(2):194–9. Epub 2011/11/19. doi: [10.1016/j.jconrel.2011.10.034](https://doi.org/10.1016/j.jconrel.2011.10.034) PMID: [22094102](https://pubmed.ncbi.nlm.nih.gov/22094102/); PubMed Central PMCID: PMC3321390.
68. Griffiths W, Crick P, Wang Y, Ogundare M, Tuschl K, Morris A, et al. Analytical strategies for characterization of oxysterol lipidomes: Liver X receptor ligands in plasma. *Free Radical Biology and Medicine*. 2013; 59:69–84. doi: [10.1016/j.freeradbiomed.2012.07.027](https://doi.org/10.1016/j.freeradbiomed.2012.07.027) WOS:000318202800008. PMID: [22846477](https://pubmed.ncbi.nlm.nih.gov/22846477/)
69. Bartel DP. MicroRNAs: target recognition and regulatory functions. *Cell*. 2009; 136(2):215–33. Epub 2009/01/27. S0092-8674(09)00008-7 [pii] doi: [10.1016/j.cell.2009.01.002](https://doi.org/10.1016/j.cell.2009.01.002) PMID: [19167326](https://pubmed.ncbi.nlm.nih.gov/19167326/).
70. Dweep H, Sticht C, Pandey P, Gretz N. miRWalk—database: prediction of possible miRNA binding sites by "walking" the genes of three genomes. *Journal of biomedical informatics*. 2011; 44(5):839–47. doi: [10.1016/j.jbi.2011.05.002](https://doi.org/10.1016/j.jbi.2011.05.002) PMID: [21605702](https://pubmed.ncbi.nlm.nih.gov/21605702/).
71. Aerts S, Van Loo P, Thijs G, Mayer H, de Martin R, Moreau Y, et al. TOUCAN 2: the all-inclusive open source workbench for regulatory sequence analysis. *Nucleic acids research*. 2005; 33(Web Server issue):W393–6. Epub 2005/06/28. doi: [10.1093/nar/gki354](https://doi.org/10.1093/nar/gki354) PMID: [15980497](https://pubmed.ncbi.nlm.nih.gov/15980497/); PubMed Central PMCID: PMC1160115.
72. Johnson DS, Mortazavi A, Myers RM, Wold B. Genome-wide mapping of in vivo protein-DNA interactions. *Science*. 2007; 316(5830):1497–502. Epub 2007/06/02. doi: [10.1126/science.1141319](https://doi.org/10.1126/science.1141319) PMID: [17540862](https://pubmed.ncbi.nlm.nih.gov/17540862/).